

Improving biological knowledge of albacore tuna, *Thunnus alalunga*, in the Indian Ocean: a scoping study

Prepared for The Food and Agriculture Organization of the United Nations

March 2020



Prepared by:

Bradley Moore¹, Adam Langley² Jessica Farley³ and Simon Hoyle¹

¹National Institute of Water and Atmospheric Research, Nelson 7010, New Zealand

For any information regarding this report please contact:

Dr Bradley Moore Fisheries Scientist

+64-3-545 7746 bradley.moore@niwa.co.nz

National Institute of Water & Atmospheric Research Ltd PO Box 893 Nelson 7040

Phone +64 3 548 1715

NIWA CLIENT REPORT No: 2020076WN Report date: March 2020 NIWA Project: FAO19401

Quality Assurance Stateme	uality Assurance Statement					
Sunfur Devis	Reviewed by:	Jennifer Devine				
AN O	Formatting checked by:	Jeanne Wissing				
R.Hur.	Approved for release by:	Rosemary Hurst				

Photo credit: <<Albacore tuna, Thunnus alalunga. [Les Hata, © SPC]

© All rights reserved. This publication may not be reproduced or copied in any form without the permission of the copyright owner(s). Such permission is only to be given in accordance with the terms of the client's contract with NIWA. This copyright extends to all forms of copying and any storage of material in any kind of information retrieval system.

Whilst NIWA has used all reasonable endeavours to ensure that the information contained in this document is accurate, NIWA does not give any express or implied warranty as to the completeness of the information contained herein, or that it will be suitable for any purpose(s) other than those specifically contemplated during the Project or agreed by NIWA and the Client.

²Fisheries Information Systems Ltd, Nelson, New Zealand

³CSIRO Oceans and Atmosphere, GPO Box 1538, Hobart, TAS 7001, Australia

Contents

Exe	cutive s	summary	6		
1	Intro	Introduction8			
	1.1	Biology of albacore tuna, with reference to the Indian Ocean	9		
	1.2	This study	10		
2		Assessing the impact of uncertainty in biological parameters on stock assessments of albacore tuna in the Indian Ocean			
	2.1	Approach	11		
	2.2	Results	12		
3	Dete	Determining sampling needs			
	3.1	Approach	14		
	3.2	Results	16		
4	Desi	Design of a biological sampling program for albacore tuna in the Indian Ocean 18			
	4.1	Goals and objectives	18		
	4.2	Biological considerations	18		
	4.3	Additional sampling considerations	20		
	4.4	Sampling strategy	28		
	4.5	Laboratory processing, including workshops and capacity development	32		
	4.6	Timeline and budget	33		
	4.7	Institutional arrangements	34		
5	Sum	mary	34		
6	Ackr	nowledgements	35		
7	Refe	rences	36		

Tables		
Table 1:	Parameters evaluated and a description of the model sensitivities investigative the influence of key biological parameters from the base Indian Ocean	ating
	albacore tuna assessment model of Langley (2019).	41
Table 2:	Change in the values of the likelihood components relative to the Base mo	odel. 42
Table 3:	Changes in individual components relative to the Base model.	43
Table 4:	Comparison of stock status and yields and associated standard deviations (derived from covariance matrix) from the range of model options.	44
Table 5:	Estimated number of hooks and sets deployed, catches and potential samp yield per assessment region for the Taiwanese longline fleet.	pling 45
Table 6:	Timeline of proposed project activities under Option A.	46
Table 7:	Indicative budget (thousand USD) for the observer-based sampling, proces analyses and reporting on Indian Ocean albacore biology under Option A.	ssing, 47
Table 8:	Timeline of proposed project activities under Option B.	48
Table 9:	Indicative budget (thousand USD) for the observer-based sampling, proces analyses and reporting on Indian Ocean albacore biology under Option B.	ssing, 49
Figures		
Figure 1:	Total annual catch (mt) of albacore tuna by fishing method from 1950 to 2 DN, driftnet; PS, purse-seine; LL, longline. Source: IOTC 2019b.	2017. 50
Figure 2:	Total annual longline catches of albacore tuna, 2012–2017.	51
Figure 3:	Total longline catch of albacore tuna by fleet, 2016–2017.	52
Figure 4:	Total annual purse-seine catches of albacore tuna, 2012–2017.	53
Figure 5:	Total purse-seine catch of albacore tuna by fleet, 2016–2017.	54
Figure 6:	Comparison of the length-weight relationship used in the base model and model sensitivity <i>LengthWeight</i> .	the 55
Figure 7:	Comparison of the growth functions used in the base model and the mode sensitivity <i>GrowthYoung</i> .	el 56
Figure 8:	Parameterisation of the variation in length-at-age from the base model and the model sensitivity <i>LengthAtAge</i> .	id 57
Figure 9:	Comparison of the growth functions used in the base model and the growt assumed for the eastern Indian Ocean in the <i>EastWestGrowth</i> model sensitivity.	th 58
Figure 10:	Spawning biomass trajectories for the base model and the base model with simulated length composition data or simulated age composition data included.	:h 59
Figure 11:	Annual CV of the estimated spawning biomass for the base model and the model with simulated length or age composition data included.	base 60
Figure 12:	Spawning biomass trajectories for the base model, the biased CPUE model the biased CPUE model with simulated length or age composition data included.	l and 61
Figure 13:	Annual recruitment for the base model, the biased CPUE model and the biased CPUE model with simulated length or age composition data included.	iased 62
Figure 14:	Frequency of growth model selection given differing sample sizes for fema (top) and male (bottom) albacore tuna.	ale 63

Figure 15:	CV of predicted size at ages 4 and 10 for female (top) and male (bottom) albacore tuna.	64
Figure 16:	Frequency of gonad index (GI, top) and fecundity (bottom) model selection given differing sample sizes for albacore tuna.	65
Figure 17:	CV of predicted gonad index (GI, top) and fecundity (bottom) at the 10th and 90th percentiles of length for albacore tuna.	d 66
Figure 18:	Weight frequency of individual albacore tuna measured from the Western Tuna and Billfish Fishery port sampling program, Australia.	67
Figure 19:	Total annual catch of albacore tuna by Chinese-flagged longline vessels, 201 2017.	2– 68
Figure 20:	Total annual catch of albacore tuna by Japanese-flagged longline vessels, 2012–2017.	69
Figure 21:	Numbers of albacore tuna measured by observers on Japanese longline vessels, 2013–2016.	70
Figure 22:	Total annual catch of albacore tuna by Korean-flagged longline vessels, 2012 2017.	<u>2</u> – 71
Figure 23:	Total annual catch of albacore tuna by Seychelles-flagged longline vessels, 2012–2017.	72
Figure 24:	Total annual catch of albacore tuna by Sri Lankan-flagged vessels, 2016 and 2017.	73
Figure 25:	Total annual catch of albacore tuna by Taiwanese-flagged longline vessels, 2012–2017.	74

Executive summary

This report presents results of a scoping study for improving biological knowledge of albacore tuna in the Indian Ocean Tuna Commission (IOTC) Area of Competence. The key objectives of this study were to: 1) undertake sensitivity analyses to evaluate the sensitivity of the stock assessment to the range of plausible values that might be estimated for key biological parameters, and potential structural changes to the assessment that might be informed by biological sampling; 2) undertake power analyses on key biological parameters to determine sampling needs; and 3) using the outcomes of 1 and 2 above, and an understanding of the fisheries, outline the sampling considerations and develop a sampling design required to provide estimates of length- and age-based population parameters to improve stock assessments of albacore tuna in the Indian Ocean (IO).

The sensitivity of the stock assessment results to alternative biological parameter values was investigated within the framework of the preliminary 2019 IO stock assessment model. A range of alternative model options were configured to evaluate the influence of key biological parameters on the stock assessment and status of albacore tuna in the IO, including alternative values for the lengthweight relationship, maturity-at-length ogive, variation in length-at-age, growth functions (emphasising growth differences in the younger age classes), and female natural mortality rates. The sensitivity of the model to spatial variation in growth rates was also considered by partitioning the model into two regions (east and west) that was also configured for fish in the eastern region having a higher growth rate than fish in the western region.

Estimates of stock status were most strongly influenced by changes related to the assumptions of growth. The alternate growth function and variation in length-at-age sensitivities both yielded considerably more pessimistic estimates of current stock status. Changes in the maturity ogive and female natural mortality influenced the magnitude of the reference biomass levels, although resulting stock status ratios were not appreciably different from the base model. Partitioning the model into two geographic regions (east and west of 75°E) yielded a considerably more optimistic estimate of current stock status and higher overall yields compared to the base model.

Simulation modelling was carried out to assess sample size requirements for growth and reproductive analyses by bootstrap resampling of the South Pacific Ocean albacore tuna dataset. The ability of the models to characterize the relationship between growth parameters and longitude improved with increasing sample size, but evidence for such a relationship was reliably identified even in the lower range of sample sizes. Similarly, for reproductive parameters, higher sample sizes improved the models' ability to identify relationships and to estimate parameters. Analyses suggest that sample sizes similar to those used to investigate biology of albacore tuna in the South Pacific Ocean (approximately 1000 individuals of each sex) will be sufficient for detecting spatial and temporal trends in biological parameters, provided sampling covers a broad geographical and temporal spread.

Owing to the biology of albacore tuna, several factors need to be considered when attempting to obtain unbiased estimates of key biological parameters, including the potential for interactive effects regarding annual, seasonal, latitudinal and longitudinal variation in growth, maturity, fecundity and the timing of spawning, as well as the potential for sex-specific differences and the need to ensure adequate numbers of small fish are sampled. In light of these considerations, to undertake a complete study on the biology of albacore tuna in the IO, sampling should ideally cover the full geographic range of the stock in the IO, with good representation across months/seasons, over at least a full year, and ideally multiple years.

The research plan provided here outlines important biological and experimental design considerations, preferred methods and sampling strategies, anticipated timelines and project budget. Sampling of Japanese and Taiwanese longline fleets via observers will be crucial for ensuring a regular and broad geographical and temporal spread of samples, and for ensuring that catch information is available. Two variations regarding time and budget are provided: a five-year study including three continuous years of sampling at a projected cost of approximately \$1,305,000 USD, or a four-year study involving two continuous years of sampling at a projected cost of approximately \$902,000 USD.

1 Introduction

Albacore tuna (*Thunnus alalunga*) are found in tropical, sub-tropical and temperate waters worldwide from approximately 50°N to 45°S. The species supports valuable commercial fisheries and important artisanal and subsistence fisheries across its range, representing an estimated 5% of the global commercial tuna catch in 2017 (SPC-OFP 2018). In the Indian Ocean (IO), the commercial catch of albacore tuna has grown substantially since commercial fishing commenced in the 1950s. Recent catches have fluctuated between ~30,000 to ~40,000 tonnes per year since a peak catch of ~43,000 tonnes in 2010 (IOTC 2016). The vast majority of landings are taken by longline (Figure 1), with the bulk of catches in recent years occurring in the waters north-east and south-east of Madagascar and south-west of Western Australia (Figure 2), primarily by Japanese- and Taiwanese-flagged vessels (Figure 3). Comparatively smaller catches are made by purse-seine, pole-and-line and gillnetting (Figure 1; Figure 4). Catches by purse-seine in 2016 and 2017 were made primarily by European Union (EU) fleets (Figure 5). Relatively large catches were obtained via driftnets in the mid-1980s and early 1990s (Figure 1) (IOTC 2019a, b).

Albacore tuna in the Indian Ocean is considered a single stock for assessment purposes. Knowledge of the stock structure, however, is uncertain. Larvae of albacore tuna are distributed in two separate and distinct zones, one in the eastern IO and the other in the western IO (Ueyanagi 1969), suggesting the existence of two distinct stocks. Based on morphometrics and DNA sequence analyses, Yeh et al. (1996) similarly proposed the existence of eastern and western IO stocks, separated at around 90°E. Other genetic studies have been largely uninformative in defining structure within the IO, with most global studies including only a single sampling location in the IO (e.g. Chow and Ushiama 1995; Montes et al. 2012). Examination of microsatellite DNA markers failed to differentiate between samples from the western IO and Atlantic Ocean (Montes et al. 2012), possibly due to some degree of mixing in the waters of South Africa. Lack of knowledge of the stock structure has long been considered, and remains, a key uncertainty in assessments of albacore tuna in the IO (Anon 1995; Langley and Hoyle 2016). Elsewhere, distinct northern hemisphere and southern hemisphere stocks are considered to occur in both the Atlantic and Pacific Oceans (Nikolic et al. 2017).

Although the migratory patterns of albacore tuna in the IO are poorly understood, the species is considered to be one of only four tuna species that are truly migratory and undertake seasonal migrations to specific feeding and spawning areas (Nikolic et al. 2017). Spawning of albacore tuna in the IO is thought to occur between 10°S and 30°S during summer, potentially in distinct areas in the eastern and western IO, and juveniles quickly move south of 30°S (Ueyanagi 1969; Dhurmeea et al. 2016; Nikolic et al. 2017). It has been suggested that juveniles and subadults do not return to the subtropics and tropics until they mature (Chen et al. 2005; Nikolic et al. 2017).

The 2016 stock assessment for Indian Ocean albacore tuna estimated that the stock was unlikely to be overfished and that it was unlikely that overfishing was occurring (Langley and Hoyle 2016). This assessment, however, used many biological parameters that were either uncertain or assumed. For example, in the absence of any published studies estimating growth using direct ageing methods in the IO, the most recent assessment used the growth curve of Chen et al. (2012) developed for the North Pacific Ocean, as the parameterisation was considered to be closer to the Indian Ocean (Langley and Hoyle 2016). Similarly, in the absence of maturity data, the maturity at age estimate of Farley et al. (2013a, 2014), calculated for South Pacific Ocean albacore tuna, was used (Langley and Hoyle 2016).

1.1 Biology of albacore tuna, with reference to the Indian Ocean

The biology of albacore tuna in the Atlantic and Pacific Oceans has been the focus of a number of studies and is generally well understood. In the Atlantic Ocean, sex-specific growth has been observed, with the average maximum length of males larger than females (Bard 1981). The sex ratio is approximately 1:1 in all length classes preceding sexual maturity, after which sex ratios become increasingly male biased with length (Bard 1981). Sexual maturity for populations in the northeast Atlantic is reached between 90 and 94 cm fork length (FL), at around 5 years of age, with no differences noted between sexes (Bard 1981).

Sex-specific and spatial variation in growth have been observed for albacore tuna in the both the South Pacific Ocean (Williams et al. 2012) and North Pacific Ocean (Chen et al. 2012). In the South Pacific Ocean, males obtain greater lengths than females for a given age from around four years of age, and individuals of both sexes obtain greater length-at-age at more easterly longitudes than at westerly longitudes (Williams et al. 2012). Longitudinal variation in length at age may also occur in the North Pacific Ocean stock (Xu et al. 2014).

In contrast to populations in the Atlantic and Pacific Oceans, there have been very few and limited studies on the biology of albacore tuna in the IO. For example, prior to 2019, there were no direct age estimates of growth available for albacore tuna anywhere in the IO. Accordingly, the IOTC Working Party on Temperate Tunas (WPTmT) recently identified the need to improve the understanding of life-history parameters for albacore tuna in the IO, and to improve understanding of the uncertainty in key demographic parameters on stock assessment results. In particular, it was noted that collaborative research across facilities on albacore tuna biology, including age and growth, length and age at maturity, and fecundity at age/length, was required, and recommended such collaborative research be undertaken as a high priority in recent WPTmT Programs of Work (IOTC 2014, 2016, 2019c), including as the top priority by the most recent WPTmT (IOTC 2019d).

Dhurmeea et al. (2016) provide an account of the reproductive biology of albacore tuna in the western IO, based on 1,790 fish (923 females and 867 males), obtained from pole-and-line, purse-seine and longline vessels. Consistent with South Pacific stocks, spawning in albacore tuna in the IO was found to occur between 10°S and 30°S, from October to January (Dhurmeea et al. 2016). Larger females were found to have a longer spawning period than smaller individuals. The mean length at 50% maturity was estimated at 85.3 cm FL. As with elsewhere, albacore tuna in the IO were found to be batch spawners, spawning on average every 2.2 days within the spawning region and spawning months. Their results, however, were considered likely biased due to a lack of small (< 65 cm FL) fish (IOTC 2019e), and were limited to only a small geographical area in the IO.

In 2017, the IOTC Secretariat developed and funded a research project to estimate the age and growth of albacore tuna in the IO. Preliminary findings of this research were provided in Farley et al. (2019). Whilst providing a preliminary examination of the growth of albacore tuna in the IO using direct ageing techniques, and of female maturity at age based on samples from Dhurmeea et al. (2016), results were limited in both time and space, being based on 600 fish collected across two years across a small geographical area (between latitudes 0–40°S and 15–65°E), and limited by a lack of small fish. Accordingly, Farley et al. (2019) make several recommendations to further this work, including that otoliths from small fish (particularly < 75 cm FL) be collected and analysed to improve growth estimations; that ovaries and otoliths from small fish (particularly < 90 cm FL) be collected and analysed to improve maturity estimations; and that otolith and ovary sampling be expanded across the IO to examine spatial patterns in growth and maturity. Farley et al. (2019) also recommended that if otoliths from fish aged < 1 year are collected, that daily ageing be undertaken on (i) the longitudinal

section to estimate length-at-age of very small fish and (ii) the transverse sections to confirm the location of the first opaque growth zone; that further work be undertaken to examine the timing of increment formation and refine the age algorithm; and that direct validation of the age estimation methods is undertaken in the IO.

1.2 This study

In 2019 the Food and Agriculture Organization of the United Nations, through the IOTC Secretariat, commissioned a scoping study to provide recommendations for improving the range of biological information for albacore tuna across the broader IO. The key objectives of the scoping study were to:

- 1. Undertake sensitivity analyses to evaluate the sensitivity of the stock assessment to:
 - 1.1 the range of plausible values that might be estimated for each parameter; and
 - 1.2 potential structural changes to the assessment that might be informed by biological sampling.
- 2. Undertake power analyses on key biological parameters to determine sampling needs.
- 3. Using the outcomes of 1 and 2 above, and an understanding of the fisheries, outline the sampling considerations and develop a sampling design required to provide estimates of length- and age-based population parameters to improve stock assessments of albacore tuna in the IO.

This report describes the results of this scoping study.

2 Assessing the impact of uncertainty in biological parameters on stock assessments of albacore tuna in the Indian Ocean

2.1 Approach

For the WPTmT07, a stock assessment has been developed for IO albacore tuna using a statistical age structured population model (Langley 2019). The assessment has been implemented in Stock Synthesis (Methot and Wentzel 2013) and includes catch, length composition data and longline CPUE indices to 2017. The primary assessment model is structured as a sex-specific (two sex), spatially aggregated model (single region) including four main longline fisheries. The longline CPUE indices from the south-western area (LL3) of the model are included as the primary abundance index from 1979–2017. Longline CPUE indices from the earlier period are not considered to represent a reliable index of stock abundance.

The longline fisheries from the southern areas are estimated to have a lower selectivity of the larger fish in the population, reflecting the lower proportion of large fish in the length composition compared to the longline fisheries in the equatorial areas. Recruitment deviates are estimated for the time period for which CPUE indices are available. There is concern regarding the reliability of the length composition data from the longline fisheries (e.g. Geehan and Hoyle 2013) and, correspondingly, these data have been assigned a low weighting in the assessment model. More details of the assessment model structure and assumptions are available in the relevant WPTmT07 document (Langley 2019).

For this study, a range of alternative model options were configured to evaluate the influence of changes to the key biological parameters on the stock assessment and status of albacore tuna in the IO (Table 1). The estimates of key stock status metrics were compared to the base model of Langley (2019) to assess the sensitivity of the results to the range of biological parameters evaluated. The current study focussed on those parameters that could be refined through further biological sampling, rather than providing a full range of model sensitivities. The final stock assessment includes additional sensitivities related to the stock-recruitment relationship and natural mortality.

The range of model sensitivities included alternative published values for the length-weight relationship for Indian Ocean albacore tuna (*LengthWeight*) (Figure 6; Table 1) and maturity-at-length from western Indian Ocean albacore tuna (*MaturityOgive*). There is uncertainty in the growth estimates for the younger age classes and an alternative growth function was configured to emphasise growth differences in the younger age classes (*GrowthYoung*) (Figure 7). There is also limited information available for the accurate determination of the variation in length-at-age. The *LengthAtAge* sensitivity incorporated a higher degree of variation in length-at-age than assumed in the base model (Figure 8).

Larger length classes of albacore tuna tend to be dominated by male fish. This may be simply a function of differential growth of older male and female fish (cf Williams et al. 2012; Farley et al. 2019), although it has also been postulated that the larger female fish may have a higher mortality rate following the onset of sexual maturity (Bard 1981; Liorzou 1989). Correspondingly, the *NatMortAge* sensitivity increased the natural mortality of female fish from 0.30 to 0.35 for the age classes 4 years and older (Table 1).

Differential growth of albacore tuna has been observed for the South Pacific with growth rates increasing eastward. To investigate the sensitivity of the model to spatial variation in growth rates, it was necessary to partition the model into two regions. Thus, the *EastWest* model was configured with two model regions (delineated at 75°E) with discrete populations (i.e. no movement between regions).

The LL3 and LL4 CPUE indices represented the primary abundance indices in the western and eastern regions, respectively. The model estimated the overall distribution of recruitment between the two regions, with the distribution of recruitment amongst the two regions also allowed to vary annually. The two-region model was then configured to include two region-specific "growth morphs" with fish in the eastern region having a higher growth rate than fish in the western region (*EastWestGrowth*) (Figure 9).

A further set of models were used to evaluate the utility of collecting length and age composition data from one of the main longline fisheries (LL3; southwestern Indian Ocean). The base assessment model was used to simulate length and age composition data from the recent period (1995–2016). No sampling error (or bias) was associated with the simulated data sets, which is an unrealistic assumption. The simulated data sets were then incorporated within separate iterations of the assessment model to compare the influence of the length and age sampling data. For each scenario, the simulated data sets were assigned a high associated weighting, reflecting the precision of the simulated data (ESS age comp 50, ESS length comp 20).

A further set of simulations were conducted that incorporated a bias in the CPUE indices for the recent period; incorporating a 2% per annum increase in the CPUE indices from 2001 to 2017. The biased model was then rerun with the simulated length and age data sets (derived from the unbiased model).

2.2 Results

The alternative growth function incorporated in the *GrowthYoung* model resulted in a considerable deterioration in the fit to both the CPUE indices and the length composition data sets (from all longline fisheries) (Table 2 and Table 3). The increased variation in length-at-age (*LengthAtAge*) improved the fit to the length composition data from the southern longline fisheries (LL3 and LL4), although there was a considerable deterioration in the fit to the LL3 CPUE indices.

Partitioning the western and eastern regions of the Indian Ocean (*EastWest*) resulted in an improvement in the fit to the LL3 CPUE indices, relative to the base model. There was also a small improvement in the fit to the length composition data from each of the longline fisheries (Table 2 and Table 3). Applying the different growth functions to the eastern region of the model (*EastWestGrowth*) improved the fit to the eastern CPUE indices (LL4) but eroded the fit to the eastern length composition data (LL2 and LL4).

Of the range of model options, the estimates of stock status were most strongly influenced by the changes related to the growth parameterisation; the *GrowthYoung* and *LengthAtAge* sensitivities both yielded considerably more pessimistic estimates of current stock status (Table 4). Changes in the maturity ogive (*MaturityOgive*) and female natural mortality (*NatMortAge*) influenced the magnitude of the reference biomass levels, although the stock status ratios were not appreciably different from the base model (Table 4). Nonetheless, the *MaturityOgive* and *NatMortAge* sensitivities estimated similar yields (*MSY*) compared to the base model.

The spatial partitioning of the model into two regions (*EastWest*) yielded a considerably more optimistic estimate of current stock status and higher overall yields compared to the base model. Changing the growth functions for the eastern partition of the stock (*EastWestGrowth*) resulted in a marginal increase in *MSY* yield relative to the *EastWest* model but did not appreciably change the estimates of current stock status (Table 4).

The biomass trajectories from the models that included the simulated length or age composition data were very similar to the base model, although the inclusion of the age data did result in a positive bias

in the level of absolute biomass (15% for the terminal year) and the estimate of current stock status (10%) relative to the base model (Figure 10). This was associated with a higher estimate of the overall level of recruitment from the model. The inclusion of the length or age data resulted in a small improvement in the precision of the estimates of the recent level of biomass corresponding to the 'data rich' period (from about 2000 onwards) (Figure 11). However, this analysis is considered exploratory and simply provides an indication of the potential scale of the influence of these two sets of data. A more thorough study should incorporate an appropriate (but unknown) magnitude of sampling variation in the derivation of the two sets of data.

The introduction of the biased CPUE indices yielded a more optimistic biomass trajectory during the recent period, relative to the base model (Figure 12). The inclusion of the simulated length data did not change the trajectory of the CPUE biased model. However, the inclusion of the age data resulted in a positive bias in the magnitude of the biomass relative to the base model that was most pronounced during the "data rich" period. This was evident in the change in the estimated level of recruitment with the inclusion of the age data (Figure 13). There was only a small deterioration in the fit to the age data between the models without (age likelihood 1.86) and with (age likelihood 2.129) bias included in the CPUE indices, suggesting that an examination of the fit to the age composition data would not provide evidence of a bias in the index. This preliminary study indicates that the age (and length) composition data are uninformative about stock size (and fishing mortality) in the context of the current stock assessment framework. Instead, these data interact strongly with the available abundance information in the parameterisation of recruitment. Clearly, considerable emphasis is required to ensure that the abundance information is sufficiently reliable to maximise the utility of the sampling data collected from the fishery.

3 Determining sampling needs

The aim of this component was to determine the sample sizes required to provide representative estimates of key biological parameters for albacore tuna in the IO. This is necessary from both a population dynamics viewpoint (understanding the biology, and how this varies spatially or temporally) and to provide inputs for the stock assessment.

3.1 Approach

Simulation modelling using the dataset for albacore tuna in the South Pacific Ocean (see Williams et al. 2012; Farley et al. 2013a,b, 2014) was undertaken to determine the sampling regime required to a) achieve target levels of precision for key biological parameters, and b) identify and estimate relationships with covariates via model selection. The original dataset included 399 fishing sets, of which 254 sets included 970 females and 335 included 928 males.

The parameters investigated included:

- Growth:
 - Length at ages 4 and 10 years, and location 200°E
- Female gonad index (GI; gonad weight / FL³ x 10⁴; Farley et al. 2013a)
 - GI at 100 cm FL
- Batch fecundity
 - Fecundity at length 100 cm FL

Simulation was carried out by bootstrap resampling of the South Pacific albacore tuna dataset. We resampled the dataset with replacement, with proportions (prop) of the available data (with nset rows) equal to 10%, 20%, 30%, 40%, 60%, 80%, 100%, and 120%. Thus, we randomly selected ns sets = $prop \times nset$, with replacement. Then within each selected set we enumerated the number of fish in the original sample (nf) and randomly resampled nf fish from the set with replacement.

Within the resampled dataset, each resampled set was allocated a new set id, so that original sets that had been sampled more than once were treated as separate sets in analyses that used set id as a random effect.

Resampling was carried out 1000 times for each value of *prop*, with each realisation of the resampled data taken to represent a new random dataset.

For the growth simulations, we fitted nine models to each resampled dataset and identified the best-fit model as the one with the lowest AIC. This 'best' model was then used to predict length for fish aged 4 and 10 located at longitude 200°E.

The nine models included all possible combinations of linear and quadratic relationships between longitude and the parameters K and L_{∞} in the logistic model.

1.
$$mod_1: len \sim L_{\infty} * (1 + e^{-K.(age-t0)})^{-1}$$

2.
$$mod_l1: len \sim (L_{\infty} + L_{\infty}2. lon) * (1 + e^{-K.(age-t0)})^{-1}$$

3.
$$mod_12: len \sim (L_{\infty} + L_{\infty} 2. lon + L_{\infty} 3. lon^2) * (1 + e^{-K.(age-t0)})^{-1}$$

```
4. \text{mod\_k1: } len \sim L_{\infty} * (1 + e^{-(K+K2.lon).(age-t0)})^{-1}
```

5.
$$\operatorname{mod_k2:} \operatorname{len} \sim L_{\infty} * \left(1 + e^{-(K+K2.lon+K3.lon^2).(age-t0)}\right)^{-1}$$

6.
$$mod_{lk1}: len \sim (L_{\infty} + L_{\infty}2. lon) * (1 + e^{-(K+K2.lon).(age-t0)})^{-1}$$

7.
$$\operatorname{mod_lk2}: len \sim (L_{\infty} + L_{\infty} 2. lon + L_{\infty} 3. lon^2) * (1 + e^{-(K + K2. lon).(age - t0)})^{-1}$$

8.
$$\operatorname{mod_lk3:} len \sim (L_{\infty} + L_{\infty}2.lon) * (1 + e^{-(K+K2.lon+K3.lon^2).(age-t0)})^{-1}$$

9.
$$\operatorname{mod_lk4}: len \sim (L_{\infty} + L_{\infty}2.lon + L_{\infty}3.lon^2) * (1 + e^{-(K+K2.lon+K3.lon^2).(age-t0)})^{-1}$$

For the female GI simulations, we fitted sixteen models to each resampled dataset and identified the best fit model as the one with the lowest AIC. This 'best' model was then used to predict GI at the 10% and 90% levels of the length distribution, in November, at the median latitude and longitude, and for the first set sampled. GI was also predicted using the best model selected for the original dataset (Farley et al 2013a).

The sixteen models included many combinations of relationships among latitude, longitude, length, and month, using thin plate regression splines s() for individual parameters, and tensor product smooths te() for interactions among parameters.

- 1. Model . *GI*∼.
- 2. Model len $GI \sim s(len)$
- 3. Model mon $GI \sim s(month)$
- 4. Model len mon $GI \sim te(len, month)$
- 5. Model lon_mon $GI \sim s(lon) + s(month)$
- 6. Model lat_mon $GI \sim s(lat) + s(month)$
- 7. Model Ion lat mon $GI \sim s(lon) + s(lat) + s(month)$
- 8. Model lon:lat_mon $GI \sim te(lon, lat) + s(month)$
- 9. Model lon_len_mon $GI \sim s(lon) + s(len) + s(month)$
- 10. Model lat_len_mon $GI \sim s(lat) + s(len) + s(month)$
- 11. Model lon_lat_len_mon $GI \sim s(lon) + s(lat) + s(len) + s(month)$
- 12. Model lon:lat_len_mon $GI \sim te(lon, lat) + s(len) + s(month)$
- 13. Model Ion len:mon $GI \sim s(lon) + te(len, month)$
- 14. Model lat_len:mon $GI \sim s(lat) + te(len, month)$
- 15. Model lon:lat_len:mon $GI \sim te(lon, lat) + te(len, month)$
- 16. Model len_lon:lat:mon $GI \sim s(len) + te(lon, lat, month)$

For the female fecundity simulations, at each iteration we fitted 22 models to the resampled dataset and identified the best fit model as the one with the lowest AIC. This 'best' model was then used to predict fecundity at the 10% and 90% levels of the length distribution, in November, at the median latitude and longitude, and for the first set sampled. Fecundity was also predicted using the best model selected for the original dataset (Farley et al 2013a).

The 22 models included a number of combinations of relationships among latitude, longitude, length, and month, using thin plate regression splines s() for individual parameters, and tensor product smooths te() for interactions among parameters.

- 1. Model $. fec \sim . + re(set)$
- 2. Model mon $fec \sim s(month)$. +re(set)
- 3. Model len $fec \sim s(len) + re(set)$
- 4. Model lon $fec \sim s(lon) + re(set)$
- 5. Model lat $fec \sim s(lat) + re(set)$
- 6. Model len_mon $fec \sim s(len) + s(mon) + re(set)$
- 7. Model len_lon $fec \sim s(len) + s(lon) + re(set)$
- 8. Model len_lat $fec \sim s(len) + s(lat) + re(set)$
- 9. Model len_mon $fec \sim s(len) + s(month) + re(set)$
- 10. Model lon_lat $fec \sim s(lon) + s(lat) + re(set)$
- 11. Model mon lat $fec \sim s(month) + s(lat) + re(set)$
- 12. Model len_lon_mon $fec \sim s(len) + s(lon) + s(month) + re(set)$
- 13. Model len_lon_lat $fec \sim s(len) + s(lon) + s(lat) + re(set)$
- 14. Model lat_lat_mon $fec \sim s(len) + s(lat) + s(month) + re(set)$
- 15. Model lat_lon_mon $fec \sim s(lat) + s(lon) + s(month) + re(set)$
- 16. Model len_lon_lat_mon $fec \sim s(len) + s(lon) + s(lat) + s(month) + re(set)$
- 17. Model len:lon $fec \sim te(len, lon) + re(set)$
- 18. Model len:mon $fec \sim te(len, month) + re(set)$
- 19. Model lon:mon $fec \sim te(lon, month) + re(set)$
- 20. Model len:lat $fec \sim te(len, lat) + re(set)$
- 21. Model lon:lat $fec \sim te(lon, lat) + re(set)$
- 22. Model lat:mon $fec \sim te(lat, month) + re(set)$

3.2 Results

In the growth analyses there were differences between males and females in the rates at which the various models were selected (Figure 14). For females, model mod_lk4 was the most commonly selected at true sample sizes or higher. However, simpler models with less spatial variation in L_{∞} were also relatively common, including mod_lk3, mod_k2, and mod_lk2. With less than 80% of the true sample sizes, model mod_k2 was the most common. For males, mod_lk4 was the most commonly selected even at 10% of the true sample sizes. However, the probability of selecting this model became steadily lower with smaller sample sizes and was less than 50% when simulated sample sizes were lower than 50% of the actual sample size.

Coefficients of variation for predicted lengths at ages 4 and 10 years decreased with increasing sample size (Figure 15). At 10% of the actual sample size the CV was 3.3% and 3.4% for females, and 2.4% and

2.5% for males. These decreased with increasing sample sizes to reach 1.1% and 1.0% for females and 0.6% for males at 100% of the actual sample sizes.

In the analyses of GI, the most commonly selected models included all four parameters latitude, longitude, length, and month (Figure 16). For fecundity, almost all the selected models included length but at higher sample sizes interactions with month, latitude and longitude were more common. Higher level interactions were not explored.

Coefficients of variation for GI decreased with increasing sample size (Figure 17). At 20% of the actual sample size, the CVs for GI at the 10th and 90th length percentiles were 26% and 27%, decreasing with increasing sample sizes to reach 21% and 20% at 100% of the actual sample sizes. The CVs for fecundity at the 10th and 90th length percentiles decreased with increasing sample size for the best models in the original study (Figure 17), from 54% and 37% at 30% of the actual sample size to 47% and 28% at 100% of the actual sample size. However, for the best models selected for each sample, the CVs for fecundity increased from 32% and 24% to reach 42% and 43% at the actual sample sizes. These CVs increased because the estimates and standard errors were predicted from the best model in each case, and with higher sample sizes the fecundity analysis selected increasingly complex models with more uncertain predictions.

4 Design of a biological sampling program for albacore tuna in the Indian Ocean

4.1 Goals and objectives

The overarching goal of proposed study is to improve stock assessment and management of albacore tuna in the Indian Ocean. The specific objectives are:

- To obtain data that will contribute to, and reduce uncertainty in growth rates and length-at-age of albacore tuna in the Indian Ocean, including spatial and temporal variation;
- To obtain data that will contribute to, and reduce uncertainty in, the maturity ogive used in stock assessment models;
- To obtain data on the spatial and seasonal variation in sex ratio;
- To obtain data on the spatial and seasonal variation in spawning frequency and location;
- To obtain data on albacore tuna fecundity, and the influence of fish size and age on batch fecundity, as well as spatial and temporal (intra-annual) effects.

Our analyses in Section 2, as well as current and previous stock assessments for albacore tuna in the IO, suggest stock assessments and resulting status determinations are sensitive to uncertainties in growth, reproduction, and spatial structure (Hoyle et al. 2014; Langley and Hoyle 2016, Langley 2019). Dedicated and structured collection of biological samples and associated data is required to resolve these uncertainties. Below, we present a roadmap towards improving understanding of biological parameters of albacore tuna in the IO, beginning with a discussion of the biological and sampling considerations, followed by sampling design and analysis considerations.

4.2 Biological considerations

4.2.1 Growth

Several factors need to be considered when attempting to obtain unbiased estimates of growth parameters for each sex/year/area strata, including:

- Sex-specific patterns of growth, including spatial variation in sex ratios, observed for albacore tuna within the IO (Farley et al. 2019) and elsewhere (e.g. North Atlantic Ocean (Bard 1981), South Pacific Ocean (Williams et al. 2012)).
- Latitudinal variation in size distribution. As discussed above, the size composition of albacore tuna in the IO varies with latitude, consistent with stocks in the Atlantic and Pacific Oceans. Generally, larger, mature albacore tuna are concentrated in equatorial areas, while smaller, immature albacore tuna are distributed at higher latitudes, with a boundary at about 30°S that roughly corresponds to the occurrence of the Circumpolar Current (Chen et al. 2005; Geehan and Hoyle 2013).
- The potential for longitudinal differences in growth, such as observed in South Pacific albacore tuna (Williams et al. 2012). Such variation may result from spatial variation in growth, selectivity, or size-dependent movement (Williams et al. 2012).

4.2.2 Reproductive parameters

Maturity

Robust, IO-wide estimates of maturity depend on sampling both immature and mature individuals in an unbiased way. Estimating the maturity schedule is difficult for species such as albacore tuna, where the mature fish migrate to discrete areas to spawn, or where there is any bias towards mature/immature fish in the sampling program. Spawning of albacore tuna in the IO is thought to occur between 10°S and 30°S during summer, potentially in distinct areas in the eastern and western IO, and juveniles quickly move south of 30°S (Ueyanagi 1969; Dhurmeea et al. 2016; Nikolic et al. 2017). It has been suggested that juveniles and subadults do not return to the subtropics and tropics until they mature (Chen et al. 2005; Nikolic et al. 2017). Important considerations include:

- Seasonal and latitudinal effects on length and age at maturity. Farley et al. (2014) observed that the proportion of mature females at length varied significantly with latitude and time of year for albacore tuna in the South Pacific Ocean. Specifically, females at northern latitudes (~10–20°S) were mature at significantly smaller lengths and ages than females at southern latitudes (~20–40°S), particularly during the spawning season, a result Farley et al. (2019) attributed to different geographic distributions of mature and immature fish during the year, and the northward movement of mature fish. Such patterns have a significant effect on maturity ogives and subsequent calculations of reproductive output, making it critical to account for such spatial and temporal variability when designing a sampling program (Farley et al. 2019). For example, biasing sampling towards lower latitudes would result in a smaller estimate of length at maturity than if fish from higher latitudes were included in the sample.
- Longitudinal effects on length and age at maturity. Given the potential for longitudinal variation in growth, longitudinal differences in maturity also need to be considered. Sampling should thus cover the full longitudinal range of albacore tuna in the IO, whilst simultaneously covering the full latitudinal range.
- Sampling time. In the South Pacific Ocean, Farley et al. (2014) reported that mature but regenerating female albacore tuna can be distinguished from immature females during the non-spawning season, and that the proportion of mature females at length varied significantly with time of year and latitude. Accordingly, year-round sampling covering all latitudes is required for estimating maturity in albacore tuna.

Sex ratio

• Information on the sex ratio of albacore tuna in the IO are largely incomplete. In the South Pacific Ocean, available data indicate that the sex ratio is slightly female-biased for smaller size classes and around 1:1 in all length classes immediately preceding sexual maturity, after which sex ratios become increasingly male-biased with length (Farley et al. 2013a).

Spawning season

■ Sampling time, and potential for longitudinal and latitudinal variation. Dhurmeea et al. (2016) observed a clear pattern of spawning of albacore tuna, with spawning concentrated between October and January and peak spawning occurring in November and December. However, their sampling was limited to the western IO (between 10°E)

and 70°E), and little is known of the timing and location of spawning in the central or eastern IO.

- Latitudinal effects also need to be considered, with Dhurmeea et al. (2016) observing spawning capable females occurring between 10°S and 30°S only during the spawning season and occurring south of 30°S outside of the spawning season (i.e., in the austral winter). Differences in spawning frequency may also be apparent: while Farley et al. (2013a) found no significant regional variation in female spawning frequency, Dhurmeea et al. (2016) observed that individuals caught from waters east of Madagascar spawned more frequently (mean spawning interval = 2.2. days) than those across the entire western IO (mean spawning interval = 3.4 days). It is recommended that monthly samples, taken from across the geographic range of albacore tuna in IO, be collected to confirm the time and location of spawning, and spawning frequency.
- Size of fish. Farley et al. (2013a) observed that the proportion of reproductively active females and the spawning fraction increased with length and age, and that larger, older fish were more reproductively active at either end of the spawning season than small, young fish; a feature that has significant implications for reproductive output. A similar pattern was found by Dhurmeea et al. (2016) for albacore tuna in the western IO, who observed a higher proportion of smaller-sized females in regressing and regenerating phases compared to large females, which were mostly in the spawning capable phase during the same period. To confirm such a pattern exists over the broader IO, a range of lengths should be sampled, with samples collected over a monthly basis across the full range of latitudes.

Fecundity and reproductive output

Potential for area differences. Farley et al. (2013a) observed that the average gonad index varied with longitude in South Pacific albacore tuna, with fish caught from easterly longitudes having heavier gonads for their size than fish caught from westerly longitudes. Similarly for the South Pacific stock, batch fecundity was found to be higher early in the spawning season (i.e., Oct-Dec) than later in the spawning season (i.e., Jan-Mar) for a given length and age, and larger, older fish had a higher batch fecundity than smaller, younger fish, in a given month (Farley et al. 2013a).

4.3 Additional sampling considerations

4.3.1 Ensuring sampling includes small fish

Obtaining small fish (i.e., < 75 cm FL) represents a significant challenge, and one that will require considerable collaborative effort and coordination to resolve. Sampling of small fish is critical for obtaining accurate assessments of growth parameters and for length and age at maturity estimations.

In the South Pacific study (Williams et al. 2012; Farley et al. 2013a,b, 2014), small fish were sourced from the troll fishery in New Zealand, and from recreational fishers along Australia's east coast, and in particular Tasmania, i.e., between approximately 40-45°S. Comparable options are generally not available in the IO. Catches by South African-flagged vessels are largely made by the pole-and-line fleet, which target juvenile and sub-adult albacore tuna between 2 and 3 years old (average of 86 cm FL), primarily in the Atlantic Ocean, with only occasional forays into the Indian Ocean (Parker et al. 2018). In South African waters, smaller catches are made by longline vessels operating with the IOTC Area of Competence, typically between 26°S and 36°S, and thus are largely comprised of adult fish

(Parker et al. 2018). According to IOTC records, less than 1% of fish from which fork length measurements exist in the IOTC database (over 4.6. million fish) are recorded as having a length of 50 cm or less, with the majority of these small individuals reported by fresh and deep-freezing Taiwanese longliners in the years between 2012 and 2017 (for FLL) and early 2000s (for LL) (IOTC 2019e).

4.3.2 Sample sizes

It is likely that a small percentage of collected otoliths will be sufficiently broken upon collection to make sectioning unsuitable, or generally unreadable. Therefore, a larger number of otoliths will need to be collected than processed.

4.3.3 Stratified vs. random sampling

Deciding whether to randomly sub-sample fish from each trip/set or conduct length stratified sampling is a topical issue in biological sampling programs. On the one hand, random sampling often leaves older individuals poorly represented in biological studies. Because these fish also tend to be larger, there is an incentive to disproportionally sample large fish. On the other, stratifying sample collection by size (length) has been shown to bias estimates of length-at-age, fitted VB parameters, and derivative estimates of natural mortality (Goodyear 2019). Mixed designs, such as supplemented sampling of large fish, have been demonstrated to similarly bias results, sometimes more than fully size-stratified designs (Goodyear 2019). Such biases may be corrected using age-length keys (ALKs), drawn from a random sample of the population (Goodyear 2019), however given uncertainties raised regarding the representativeness of length composition data for albacore tuna in the IO (Geehan and Hoyle 2013), the use of an ALK for this purpose is not recommended. Accordingly, it is recommended that random sampling of fish within sets be undertaken.

4.3.4 Fleet considerations

A number of nations have fished for albacore tuna in the IOTC Area of Competence in recent years. Below we provide a synthesis of the main fishing nations, focusing on recent catch statistics, how fish are processed, and current observer and port sampling programs.

Australia: The number of active Australian-flagged longline vessels and levels of fishing effort in the IO have remained low in recent years, largely due to reduced profitability. In 2017, three vessels from the Western Tuna and Billfish Fishery (WTBF) and seven vessels from the Eastern Tuna and Billfish Fishery (ETBF) fished in the IOTC Area of Competence (Hobsbawn et al. 2018). The total catch of albacore tuna by all Australian flagged longline vessels in the IOTC Area of Competence was 18.6 t (Hobsbawn et al. 2018), equating to around 890 individuals. Most fish are landed fresh (on ice or brine chilled) (K. Williams, WW Fisheries, pers. comm.). A further 0.2 t was caught by Commonwealth multipurpose vessels (trolling, pole-and-line, dropline and handline). No albacore tuna were reported in Western Australian state fisheries catches in 2016 or 2017. Estimates of the recreational catch for tuna, including albacore tuna, within the IOTC Area of Competence in Australian waters is uncertain (Hobsbawn et al. 2018).

In 2007, an observer program was implemented in the WTBF. Since 1 July 2015, all observer coverage on Australian-flagged vessels in the WTBF has been via electronic monitoring (Hobsbawn et al. 2018). A port sampling program for the WTBF has been in place since 1999, with weight data for individual fish collected by a single operator (of 2 vessels) based in Freemantle (K. Williams, pers. comm.). In 2017, 854 individual albacore tuna were measured within the IOTC Area of Competence by this program (Hobsbawn et al. 2018). Most fish caught are between 10 and 15 kg ($^{\sim}$ 78–90 cm FL), however fish < 5 kg (and thus likely less than 60 cm FL based on the length-weight relationship of Williams et al. 2012) are caught occasionally (Figure 18). However, the lack of 'on-the-ground' staff may mean

sampling personnel have to travel to this location on a regular basis, making monthly sampling difficult and/or costly.

China: The Chinese-flagged longline fleet operating in the IOTC Area of Competence consisted of 71 deep-frozen longliners and 10 fresh longliners in 2017 (Zhu et al. 2018). Combined, these vessels caught 3,646 mt of albacore tuna in 2017, with 1,320 mt taken by deep-frozen longliners and 2,326 mt taken by fresh longliners. In 2017, catches from both fleets predominantly came from the southern IO (Zhu et al. 2018; Figure 19).

China has operated an observer program in the IOTC Area of Competence since 2002. Four observers were deployed in 2017, all on deep-frozen longliners, with observer trips occurring in the north-west IO, immediately east and south-east of Madagascar, and the southern IO between approximately 50°E and 80°E at around 33°S (Jiangfeng Zhu, Shanghai Ocean University, pers. comm., Zhu et al. 2018). The total number of albacore tuna measured through this program is low (e.g. 52 individual fish in 2012, 26 in 2013; Zou et al. 2014), although observer coverage has increased in recent years. China has also implemented a port sampling program since 2012. In 2017, 300 albacore tuna were measured in this sampling program, though a key challenge of this sampling is the lack of detailed capture information, particularly around catch date and location (Zhu et al. 2018).

Comoros: The Comoros fleet operating in the IOTC Area of Competence is exclusively artisanal, with vessels ranging up to approximately 25 miles from the coast, with durations of up to a week. Fisheries use a variety of gears, in particular handline, trolling and vertical longline / dropstone, and land fish fresh. The fleet caught an estimated 37 t of albacore tuna in 2017. The artisanal nature of the fleet and vast number of geographically disparate landing sites makes sampling at this location difficult.

EU-France: Excluding the overseas departments of Réunion and Mayotte (discussed separately below), the French tuna fishing fleet operating in the IOTC Area of Competence in 2017 consisted of 12 purse-seine vessels (Bach et al. 2018). Landings are mainly in Victoria (Seychelles), Port Louis (Mauritius) and Diego Suarez (Madagascar). Combined, these vessels caught 66,945 t of tuna in 2017, with albacore tuna constituting less than 0.5% of the total catch (149 mt) (Bach et al. 2018).

EU-France-Mayotte: The Mahoran longline fleet consisted of three active vessels in 2017, all of which targeted swordfish in surface waters (Bach et al. 2018). Catches of albacore tuna by this fleet are unknown but likely to be negligible, with a total of 30.6 t of tuna (all species) caught in 2017, with the bulk of this being yellowfin tuna (*Thunnus albacares*) and bigeye tuna (*Thunnus obesus*) (Bach et al. 2018).

An extensive small-scale fleet operates in coastal and nearshore waters of Mayotte, consisting of 141 registered 'professional' vessels in 2017, along with around 300 un-registered 'non-professional' vessels that use similar gears, and around 700 canoes. In 2017, the registered portion of this fleet caught 646 t of pelagic and demersal fish (Bach et al. 2018). Catches of albacore tuna by this fleet are unknown but considered negligible.

EU-France-Reunion: There were 40 longline vessels operating out of Réunion in 2018, 19 of which were 'semi-industrial' (12–24m in length) vessels that operated in offshore waters and 21 of which were smaller vessels that primarily operated in coastal waters (S. Bonhommeau, IFREMER, pers. comm.). In addition, a number (131 in 2018) of small-scale fishing boats that use handline and small longlines occasionally catch albacore tuna in the nearshore waters around Réunion. The offshore longline fleet caught 195 mt of albacore tuna in 2018 and approximately 149 mt of albacore tuna in 2017, which constituted around 13% of the total catch of this fleet (S. Bonhommeau, pers. comm., IOTC 2019a). The coastal longline fleet caught approximately 65 mt of albacore tuna in 2018 (equating

to approximately 16% of the total catch and 37% of the tuna catch of this fleet), while the small-scale fleet caught approximately 18.7 mt in 2018 (equating to approximately 2% of the total catch of this fleet and 6% of the fleet's tuna catch) (S. Bonhommeau, pers. comm.). All catches are landed in Réunion, with the majority of catches landed fresh.

An observer program for the offshore longline fleet has been operation since 2010; the observer program covers approximately 10% of trips (S. Bonhommeau, pers. comm.).

Réunion has a port sampling program in place for its semi-industrial and coastal longline fleets. In 2018, 590 albacore tuna were measured through this sampling program, including 510 from the semi-industrial fleet and 80 from the small coastal longline fleet (S. Bonhommeau, pers. comm.). In addition, there is an established, ongoing biological sampling program in Réunion, with approximately 150 albacore tuna sampled for otoliths, gonads and muscle each year since 2018 (ranging from around 90–110 cm FL) (S. Bonhommeau, pers. comm.). Fish are generally gutted upon landing at Réunion, but fishers are compensated to keep the viscera for a limited number of fish. Most fish are sold whole (i.e. with head intact), but a small number are sold without the head, and can thus be sampled for otoliths (S. Bonhommeau, pers. comm.).

EU-Italy: The Italian fishing fleet operating in the IOTC Area of Competence in 2017 consisted of a single purse-seine vessel. This vessel caught a total of 2 mt of albacore tuna in 2017 (UE-Italy 2018).

EU-Portugal: The Portuguese fishing vessels operating in the IOTC Area of Competence consisted only of pelagic longliners targeting swordfish (Coelho 2018). Catches of albacore tuna by this fleet are negligible, with < 6 mt caught in 2017 (IOTC 2019a).

EU-Spain: There were 14 EU-Spanish-flagged industrial longline vessels operating in the IOTC Area of Competence in 2017, along with 14 purse-seine vessels. Spanish-flagged longline vessels do not target albacore tuna, and have not reported any albacore tuna catches in recent years.

The purse-seine fleet caught 100 t of albacore tuna in 2017 (0.07% of the fleets' total annual purse-seine catch for that year), largely as by-catch (IEO and SGP 2018).

EU-United Kingdom: The UK fishing fleet operating in the IOTC Area of Competence in 2017 consisted of two longline vessels. These vessels mainly target swordfish and sharks, and caught a total of 579.8 mt in the IOTC Area of Competence in 2017, including 3.1 mt of albacore tuna (constituting approximately 0.5% of the total catch) (United Kingdom 2018).

India: Little is known of albacore tuna catches by Indian-flagged vessels. Catches are likely to be negligible, with no reference to albacore tuna made in the most recent submitted national report to the IOTC Scientific Committee (Premchand et al. 2015), and no catches of albacore tuna reported in data submitted to the IOTC (IOTC 2019a).

Indonesia: Indonesian-flagged vessels caught 6,994 t of albacore tuna in the IO in 2017, with the majority (6,399 t) landed by longline vessels (Ruchimat et al. 2018). Approximately 357 t of albacore tuna were landed at Benoa Port in 2017 (Ruchimat et al. 2018), where staff from the Research Institute for Tuna Fisheries undertake port sampling. In general, larger sized individuals are landed (i.e., >90 cm FL), with no apparent seasonality in catches (Ririk Sulistyaningsih, Research Institute for Tuna Fisheries (RITF), pers. comm.). However, most of the albacore tuna landed in Benoa are frozen without processing and sold whole, with vendors opposed to any invasive sampling resulting in damage to the fish (Ririk Sulistyaningsih, pers. comm.) as they are cooked whole for canning, precluding the acquisition of biological samples such as otolith and gonads from these fish. Samples may be obtained

by purchasing fish, with an individual fish costing approximately \$50–\$60 USD at the time of writing (Ririk Sulistyaningsih, pers. comm.).

Fresh albacore tuna caught in the IOTC Area of Competence are landed at the ports of Sendang Biru and Pacitan on the island of Java. In 2016, landings were around 500 mt and 213 mt, respectively (Ririk Sulistyaningsih, pers. comm.). Landings at these ports are more seasonal than in Benoa, with the majority of fish landed between May and August (Ririk Sulistyaningsih, pers. comm.). Fish are landed and sold whole for canning at Sedang and landed gilled and gutted at Pacitan. There is no existing sampling program for these ports, and sampling would require translocating a sampling team from Bali or Jakarta (and thus monthly sampling may prove difficult and/or costly). However, RITF expects that there should be both enumerators and observers based at these ports by 2020, that may assist with sampling (Ririk Sulistyaningsih, pers. comm.).

Islamic Republic of Iran: Little is known of albacore tuna catches by Iranian vessels, with no reference to albacore tuna in the recent national report to the Scientific Committee of the IOTC (Anon 2018a). Catches are likely to be low, with no industrial longline vessels in operation (Anon 2018a). According to the IOTC, offshore gillnet vessels from I.R. Iran have extended their area of operation in recent years, and are now thought to operate on the high seas closer to the equator. However, the lack of catch-and-effort data from these fleets makes it difficult to assess whether they are operating in areas where catches of juvenile albacore tuna are likely to occur (IOTC 2019e).

Japan: Japan had 41 longliners operating in the IOTC Area of Competence in 2017, along with 3 purse-seiners. The longline fleet caught a total of 1,668 mt in 2017, a decline of 699 mt from the previous year (Anon 2018b). Since the 1960s, the species was largely caught as by-catch, although in recent years more Japanese longline vessels have been targeting albacore tuna, especially in the waters south-west of Australia and, to a lesser extent, around South Africa (Figure 20), largely due to decreased quota of southern bluefin tuna and enhanced market value of albacore tuna for sashimi products (Matsumoto 2016). No albacore tuna catch was reported from the purse-seine fleet in 2016 or 2017, consistent with the long-term trend of this fishery. The longline catch is unloaded abroad predominantly in a frozen state (Anon 2018b).

Japan has operated an observer program in the IOTC Area of Competence since 2010. During 2010–2016, observer numbers have ranged between 6 and 14 (average of 10) per year, covering an average of 6.6% of hooks annually (Anon 2018b). In 2016, observers measured a total of 3,658 individual albacore tuna from a total of 9 vessels (Anon 2018b). In recent years, the bulk of observer coverage has been concentrated around the waters south-west of Australia (Figure 21), consistent with patterns in catch.

Kenya: Approximately 1,931 t of tuna (all species) were landed by artisanal fisheries in 2017 (Mueni et al. 2018). Due to the artisanal nature of the fleet it is not possible to identity the proportional of albacore tuna in the catch, though it is considered to be negligible. No port sampling of artisanal vessels is currently conducted (Mueni et al. 2018).

Madagascar: The Malagasy fleet operating in the IOTC Area of Competence in 2017 was comprised of seven longliners all less than 24 m in length. Since 2014, these longliners have operated exclusively in the waters east of Madagascar, generally been 14°S and 22°S. In 2017, the fleet caught approximately 39 t of albacore tuna within the IOTC Area of Competence, which was approximately half of the catch for 2016 (79 t). Port sampling is conducted in two ports on the east coast – Sainte Marie and Tamatave – where two and five longliners of the national fleet are based, respectively (MRHP et al. 2018).

Malaysia: Malaysia had 19 longline vessels operating in the IOTC Area of Competence in 2017, with 6 vessels operating in the southwest IO primarily targeting albacore tuna, and 13 operating in the east IO primarily targeting yellowfin and bigeye tunas (Samsudin et al. 2018). Combined, the vessels caught approximately 1,607 t of albacore tuna in 2017. The six vessels targeting albacore tuna unloaded their catch in a frozen state at Port Louis, Mauritius (Samsudin et al. 2018), precluding port sampling of this catch. At the time of writing, Malaysia did not have a national observer scheme, precluding any onboard sampling at present, although were looking to implement an observer program in mid-2019 (Samsudin et al. 2018).

Mauritius: Mauritius had 12 semi-industrial longliners operating in the IOTC Area of Competence in 2017, along with 2 purse-seiners (and one supply vessel). The longline vessels typically carry out short trips (9–11 days) and land most of their fish chilled. The total catch of albacore tuna by the semi-industrial longline fleet in 2017 was 36 mt, with the majority of this (34 mt) taken by longliners operating within the Mauritius EEZ, and the remainder taken off the coast of, and landed in, Mozambique (Sheik Mamode et al. 2018; Clivy Lim Shung, Ministry of Ocean Economy, Marine Resources, Fisheries and Shipping, pers. comm.). Small amounts (around 1 mt) of albacore tuna were taken by the purse-seine fishery in 2017 (Sheik Mamode et al. 2018). The semi-industrial longline fleet is currently not covered by human observers. Observers are deployed on one of the purse-seine vessels (Sheik Mamode et al. 2018). Some port sampling of the semi-industrial longline catch is conducted at Port Louis, with 195 individual albacore tuna measured through this program to date (Sheik Mamode et al. 2018).

Mozambique: Mozambique had 2 industrial longliners licenced in 2017. The total catch of albacore tuna in 2017 by the national fleet was 0.84 t (Chacate and Mutombene 2018), equating to approximately 35 individuals. No albacore tuna were reported in artisanal catches in either 2016 or 2017 (Chacate and Mutombene 2018).

Oman: Little is known of albacore tuna catches by Omani-flagged vessels, with no reference to albacore tuna in the recent national report to the Scientific Committee of the IOTC (MAF 2018). Catches are likely to be low, with only one industrial longline vessel in operation (MAF 2018).

Pakistan: Little is known of albacore tuna catches by Pakistan vessels, with no reference to albacore tuna in the recent national report to the Scientific Committee of the IOTC (Khan 2018). Catches are likely to be low, with no industrial longline vessels in operation, and the majority of vessels using gillnets to catch yellowfin tuna, neritic tunas such as longtail tuna (*Thunnus tonggol*) and frigate tuna (*Auxis thazard*), and other nearshore pelagics (Khan 2018). According to the IOTC, offshore gillnet vessels from Pakistan have extended their area of operation in recent years, and are now thought to operate on the high seas closer to the equator. However, the lack of catch-and-effort data from these fleets makes it difficult to assess whether they are operating in areas where catches of juvenile albacore tuna are likely to occur (IOTC 2019e).

Philippines: The Philippines had only one active vessel operating in the IOTC Area of Competence in 2017, a purse-seiner. No albacore tuna were reported in catches from this vessel (Gongona et al. 2018).

Rep. of Korea: There were 13 Korean-flagged longline vessels operating in the IOTC Area of Competence in 2017, along with 3 purse-seine vessels. The longline fleet caught approximately 6,625 individual albacore tuna in 2017, down from 9,640 in 2016 and 16,656 in 2015 (Kim et al. 2018). This decline was largely related to a shift in fishing location, with a number of vessels shifting to target

yellowfin tuna and bigeye tuna in the western tropical area between 5°N and 10°S and in the Mozambique Channel (Kim et al. 2018; Figure 22).

Korea has operated an observer program for distant water fisheries since 2010. In 2017, two observers were dispatched on Korean-flagged longline vessels within the IOTC Area of Competence, with both observers operating on trips within the Mozambique Channel (Kim et al. 2018).

Rep. of Maldives: There were 44 Maldivian-flagged longline vessels operating in the IOTC Area of Competence in 2017, including 7 vessels > 22.5 m, along with a number of pole-and-line and troll vessels. The total catch (i.e., all species combined) of the longline fleet in 2017 was 139,000 t (Ahusan et al. 2018). Catches of albacore tuna however are likely to be negligible. Data supplied to the IOTC for the current stock assessment indicate that no albacore tuna were caught by the Maldivian-flagged industrial longline fleet in 2017 and < 5 t in 2016, with only small amounts caught by the coastal longline fleet (3.38 t in 2017) (IOTC 2019a).

Rep. of South Africa: The majority of catches of albacore tuna in South Africa are made in Atlantic Ocean waters, largely by the pole-and-line fleet operating out of Cape Town. There were 16 South African-flagged longline vessels and a single pole-and-line vessel operating in the IOTC Area of Competence in 2017, with the latter fishing in the Indian Ocean for a total of 12 hours (Parker et al. 2018). The total catch of albacore tuna by the South African large pelagic longline fleet in the IOTC Area of Competence was 26.5 t in 2017, slightly up from 19.9 t in 2016, but well below the annual catch of 2013 (177.5 t) (Parker et al. 2018). While the size structure of the catch was unknown at the time of writing, catches by the longline fleet in the IOTC Area of Competence typically occur between 26°S and 36°S (Parker et al. 2018), and thus likely to be comprised of adult fish.

One hundred percent (100%) observer coverage is required on joint venture (foreign-flagged) longline vessels, with four observers actively observing on the three Japanese joint-venture vessels operating in the IOTC region in 2017, and a single trip from a local longline vessel observed. Length frequency data from the pelagic longline fleet are collected at sea by observers prior to the fish being dressed (Parker et al. 2018). Observers collect biological material when required (Parker et al. 2018). Approximately 655 individual albacore tuna were measured by observed on pelagic longline vessels in the IOTC Area of Competence in 2017 (Parker et al. 2018). There are no observers stationed on poleand-line vessels (Parker et al. 2018).

South Africa had an active port sampling for albacore tuna (and other species), with length-frequency data collected through sampling of pole-and-line vessels by Department of Agriculture, Forestry and Fisheries (DAFF) staff from 2011–2018. The port sampling program is no longer operating due to staffing issues (D. Parker, Department of Agriculture, Forestry and Fisheries, pers. comm.). However, some port sampling is conducted on an ad-hoc basis when samples are required. Small fish (down to 60 cm FL) are occasionally available through this sampling (D. Parker, pers. comm.).

The boat-based recreational fishery also targets albacore tuna, along with other tunas and marlins. Opportunities exist to sample this fishery, such as has been previously conducted by DAFF staff in collaboration with the University of Cape Town, although this would be on an opportunistic basis (e.g. at fishing competitions). At the time of writing, the size structure of fish sampled through these activities was unknown. It is likely some small fish are caught by recreational fishers, although arrangements would have to be made to retain these. Most of the recreational catch is taken from the International Commission for the Conservation of Atlantic Tunas (ICCAT) Area of Competence (D. Parker, pers. comm.), however evidence suggests these fish may form the same stock as found in South African waters in the IOTC Area of Competence (Montes et al. 2012).

Seychelles: There were 48 Seychellois-flagged industrial longline vessels operating in the IOTC Area of Competence in 2017, along with 13 purse-seine vessels and 31 small-scale semi-industrial longline vessels (Assan et al. 2018). Data supplied to the IOTC for the current stock assessment indicate that the industrial longline fleet caught approximately 656 mt of albacore tuna in 2017, or approximately 49,947 individuals, with the bulk of the catch occurring in the western IO between 12.5°N and 37.5°S (Figure 23), by frozen longline vessels (IOTC 2019a). The purse-seine fleet caught 56 mt of albacore tuna in 2017 (Assan et al. 2018). Albacore tuna are seldom if at all caught by the small-scale longline fleet (N. Bodin, Seychelles Fisheries Authority, pers. comm.). The industrial longline fleet is currently not covered by human observers, though electronic monitoring is being trialled on two vessels (Assan et al. 2018). The industrial longline fleet does not land in Port Victoria and there are no port sampling programmes for these vessels (Assan et al. 2018). Rather, a self-sampling programme is being implemented, whereby size frequency data are being recorded by the crew and transmitted to the Seychelles Fishing Authority (Assan et al. 2018). Port sampling is a routine activity for the purse-seine, small-scale longline and local artisanal fleets, although these activities are primarily focussed on yellowfin, skipjack (Katsuwonus pelamis) and bigeye tunas (being the dominant species of these catches), with only one individual albacore tuna measured in 2017 (Assan et al. 2018).

Somalia: Catches of albacore tuna by Somali-flagged vessels are unknown, but likely to be negligible. There were no Somali-flagged vessels > 24 m operating in the IOTC Area of Competence in 2018, and albacore tuna is not a target species for artisanal fishers (Sheik Heile et al. 2018).

Sri Lanka: There were two Sri Lankan-flagged large (> 24 m) longline vessels in operation in the IOTC Area of Competence in 2017, along with a number of smaller longline, gillnet, and ring net vessels (Hewapathirana et al. 2018). The longline fleet landed an estimated 34.8 mt of albacore tuna, primarily in the northern IO, including the southern Arabian Sea and Bay of Bengal (Hewapathirana et al. 2018, IOTC 2019a). A further 33.5 mt was caught by offshore ring net, and 54.7 mt caught by offshore gillnet, in 2017 (Figure 24).

The Department of Fisheries and National Aquatic Resources Research and Development Agency (NARA) undertake port sampling in 13 of the 15 coastal fisheries districts, focusing on offshore multiday boats and tuna-targeting coastal day boats (Hewapathirana et al. 2018). However, only a small number of albacore tuna are encountered through this sampling (albacore tuna represented approximately 0.6% of all fish measured in port sampling in 2017) (Hewapathirana et al. 2018).

Taiwan, China: Taiwanese longliners have operated in the IOTC Area of Competence since the 1950s. In 2017, the Taiwanese longline fleet consisted of 138 large-scale vessels (> 100 GT) and 180 small-scale vessels (< 100 GT) (FA and OFDC 2018). In the recent decade, the catch of albacore tuna by Taiwanese longliners in the IOTC Area of Competence has fluctuated between 10,000 mt to 18,000 mt (Chang et al. 2016). In recent years longline operations have covered the geographic breadth of the IO, with the bulk of catches coming from the south western and south eastern regions (Wang 2019; Figure 3; Figure 25). The large-scale longline fleet caught approximately 5,199 mt of albacore tuna in 2017, predominantly from 30°S to 40°S, while the small-scale longline fleet caught approximately 17,283 mt, predominantly in the western IO (FA and OFDC 2018). Catches of the large-scale fleet are unloaded at Mauritius, although most vessels tranship their catch at sea (Ren-Fen Wu, Overseas Fisheries Development Council, pers. comm.), potentially limiting the catch information available. The small-scale fishery operates as two sub-fleets, with one sub-fleet targeting albacore tuna in the western IO and using Mauritius as a base (unloading at this location), and the other largely targeting tropical tuna in the eastern IO using Phuket (Thailand) and Sri Lanka as bases (unloading at these

locations). Most of the small-scale longliners have freezers on board and thus store their catches frozen (Ren-Fen Wu, pers. comm.).

Taiwan maintains an active observer program in the IOTC Area of Competence. In 2017, there were 1,894 fishing days observed by 19 observers deployed on the large-scale tuna longline vessels. For the small-scale longline fishery, there were 756 fishing days observed by 12 observers. Observers collect fisheries data and size measurements of target species and record bycatch. At the time of writing, it was unknown whether buyers would allow biological sampling of the catch.

Tanzania: No national report was available for review for Tanzania in 2017 or 2018. In 2015, Tanzania's national fleet in operation in the IOTC Area of Competence consisted of 3 commercial longliners and a number of small boats using small-scale gillnets, troll lines and longlines (Amir and Hamid 2016). The commercial longline fleet caught approximately 28 t of albacore tuna in 2015, with the bulk of catches made within the Tanzanian EEZ and in the waters north and northeast of Madagascar (Amir and Hamid 2016).

Thailand: Little is known of albacore tuna catches by Thai-flagged vessels in the IOTC Area of Competence, with no catch statistics for albacore tuna available in the recent national report to the Scientific Committee of the IOTC (Lirdwitayaprasit et al. 2018). Catches are likely to be low, with no industrial longline vessels and only a single purse-seine vessel in operation in 2016 and 2017 (Lirdwitayaprasit et al. 2018). Several foreign fleets unloaded in the Thai port of Phuket in 2017, including Bahamas, Malaysia, Panama, Taiwan, China, Republic of Korea, although these unloadings seldom have corresponding catch information available (Panjarat et al. 2016).

4.4 Sampling strategy

To sustain an effective biological sampling for albacore tuna in the IO, two important general considerations are:

- 1) sample suitability, i.e., 'fit for purpose' ensuring samples and associated information are adequate for addressing the research questions being asked, and
- 2) sampling efficiency i.e., ensuring value for effort.

In light of the considerations described above, to undertake a complete study on the biology of albacore tuna in the IO, including, age, growth and reproduction, sampling should ideally cover the full geographic range of the stock in the IO, with good representation across months/seasons, over at least a full year. Sampling should be maintained in a second year, and ideally longer (i.e. three or more years), to determine if there is variation in parameters such as growth, maturity, and sex ratios over time.

4.4.1 Fleets and approach

To obtain the requisite number of samples and spatial coverage in a manner as efficient as possible, we recommend that sampling be conducted from the following fleets:

- Distant water fishing nations, in particular Japanese and Taiwanese-flagged longliners, and potentially Chinese and Korean longliners, via observers;
- Local fleets: South African longline fleet, Réunion longline fleet (including both offshore and coastal sectors), and Mauritius semi-industrial longline fleet; potentially Tanzanian, Sri Lankan, Indonesian and Australian WTBF longline fleets.

Sampling of the Japanese and Taiwanese longline fleets via observers will be critical to ensuring a regular and broad geographical spread of samples, and ensuring that catch information (location, date, time and state of fish when landed). Moreover, each of these fisheries holds its own intrinsic value for sampling. For example, sampling from the Japanese longline fleet will be crucial for obtaining fish from the south-eastern IO, while sampling from the Taiwanese longline fleets will be crucial to ensuring broad geographical coverage, particularly in the southern latitudes.

Sampling of local fleets may provide the opportunity for supplementing samples from DWFN fleets in specific geographic areas, and offers an alternative approach to sampling in the event that sampling from DWFN fleets yield insufficient samples. However, due to staffing limitations, contractors would have to be engaged to sample these locations, increasing project costs, hence our preference for sampling via observers.

Based on our experience conducting biological studies from vessel and port sampling, operators generally prefer to sample a fixed, smaller number of fish from every set or trip rather than a variable (e.g. over time and space), larger number of fish from a subset of trips. It is also practically easier to implement a sampling strategy in which a smaller, fixed number of individuals are sampled than it is to implement a more complex protocol that requires keeping track of which trips have been sampled, especially if this is to vary in time and space. This is particularly an issue for DFWN fleets, where vessels and observers may be at sea with limited contact for many months.

The sampling needs analyses presented in Section 3 indicate that sample sizes similar to those used in the South Pacific study (approximately 1000 individuals of each sex) will be sufficient for detecting spatial and temporal trends, if present, provided sampling covers a broad geographical and temporal spread. A preliminary analysis based on Taiwanese longline data for 2017 provided by the IOTC (IOTC 2019a) indicates that a simple sampling strategy of sampling five fish from 2% of sets in a year would result in approximately 1,629 individual albacore tuna being sampled (Table 5). It should be noted that these analyses are based on assumed numbers of hooks per set (and the assumption that this is constant among regions), nominal (rather than regionally standardised) CPUE, assumed rates of zero catch set per region (adapted from Hoyle et al. 2015) and observer coverage rates that approximate those of the small-scale longline fishery (i.e., 2%, FA and OFDC 2018). These analyses indicate that such a sampling strategy would yield sufficient sample sizes in all regions except for Region 2, highlighting the importance of sampling from additional fleets, and in particular the Japanese longline fleet (Table 5; Figure 20).

Accordingly, we recommend sampling be conducted in the following manner:

- A maximum of five albacore tuna be sampled per set from Taiwanese and Japanese longline vessels. The first five fish should be set aside for sampling, to ensure samples are randomly selected from the catch per set.
- In addition, each fish smaller than 60 cm FL or greater than 110 cm FL should be retained for sampling. These fish should be considered additional to the above and marked as such on the corresponding data collection form.
- The sample size and sampling protocol should be reviewed 12 months following full implementation to assess whether the number of fish sampled per set is appropriate and whether the size of samples is unbiased.

4.4.2 What tissue to sample

To address the considerations outlined above, the following biological material and data are required:

- Sagittal otoliths (for direct age estimation);
- Whole gonads (for estimation of maturity, batch fecundity, spawning frequency/fraction, and overall reproductive output);
- First dorsal fin spine (to compare/verify age against that from otoliths; particularly for small fish i.e., < 70 cm FL);
- Fork length (to nearest cm);
- Weight, to the nearest 0.1 kg (whole and dressed for conversion factors);
- Capture date, location (longitude and latitude) and time (if possible);
- Vessel name and flag;
- Whether the fish was alive or dead when landed (this being particularly important for mature fish during the spawning season to assess the time it takes to resorb postovulatory follicles for spawning frequency calculations); and
- Fishing method and set information (e.g. hook depth, hook number (position between float), line length).

Additionally, the following material should be collected if opportunities exist (these materials are not requisite for addressing the considerations outlined above, but provide significant value-adding to a biological sampling program by facilitating examinations of population structure and genetic connectivity, short-term residency patterns, diets and food web positioning, and methylmercury and other organometallic toxin concentrations):

- Liver (a piece approximately 4–5 cm long);
- Stomach (cut as close to the gills as possible); and
- Muscle tissue (approximately 4-5 cm sample cut from the back of the fish or from near the anus).

4.4.3 Sampler training and equipment

Sampling over a broad spatial area, with temporal repetition, will accordingly require an intensive and coordinated effort, involving staff from many different Collaborating Contracting Parties and Non-Contracting Parties (CPCs), regional scientific authorities, and academic institutions. The collection of such a large dataset will also require participation by a range of experts in the analysis of samples and resulting data.

To minimise the possibility of introducing artificial variation between samples as a result of variability in sample collection, handling, management and storage methods between samplers (individuals, teams or organisations), consistent best-practice approaches should be employed. Training should be provided to all samplers prior to commencing sampling. This should include the development of appropriate training material, including theoretical modules and hands-on, practical training for collection and handling of samples as well as data collections for standardisation, similar to the Pacific Islands Regional Fisheries Observers (PIRFO) biological sampling training program. Protocols should be developed that clearly set out the objectives of the project, facilitate standardisation where possible, support the generation of useful metadata streams and provide team members with the tools they need for achieving a successful sampling programme. For sampling of albacore tuna, this

should include protocols for obtaining fish, species identification, standardised approaches for collecting tissues, sample labelling, preserving and packaging, and metadata collection standards, as well as transport and logistical arrangements (including permitting). Special attention should be given to the sample storage methods and best practices should be developed to ensure that sample quality is maintained on the long-term.

Observer coordinating agencies within CPCs should be provided with biological sampling kits. Each observer tasked to undertake biological sampling should be provided with a kit, as well as spare equipment, particularly if at sea for an extended period. A sampling kit (to sample 50 fish) should consist of:

- Knives (and sharpener);
- 1 saw;
- 1 pair fine tweezers (to extract otoliths);
- 1 large plastic bag (to store samples onboard);
- 100 large clickseal / ziplock bags (50 bags for stomachs, 50 for gonads);
- 100 medium clickseal bags / ziplock bags (50 for muscle, 50 for livers);
- 50 cable tie labels (to assist with correctly labelling samples);
- 50 small vials for otoliths;
- Pencils pencil sharpeners, erasers;
- Gel ice packs;
- Sampling instructions;
- Biological sampling pamphlet;
- Biological sampling forms;
- Small kitchen scale for weighing gonads at sea (if subsampling of gonads at sea is required); and
- Set of callipers for measuring whole fish.

4.4.4 Sample preservation and storage

Otsu and Uchida (1959) found no significant difference in the mean diameter of the most advanced group of oocytes (MAGO), or in oocyte size frequency distributions, between left and right ovaries or along the length of the ovary. However, they did find a significant difference in mean diameter of the MAGO between the periphery and centre of the ovary suggesting that oocyte development can vary across the ovary. Accordingly, should sub-sampling of the gonad be required (e.g. due to space considerations onboard fishing vessels), a core subsample (lumen to the periphery) should be taken from the mid-section of either ovary. In these instances, the weight of the whole gonad should be obtained. Ideally, samples should be fixed fresh in 10% buffered formalin for later histological sectioning. Should this not be possible (e.g. due to issues with obtaining formalin, or storage of formalin on fishing vessels), samples should be frozen immediately upon collection. A comparison of histological sections prepared from fresh and frozen-thawed material by Farley and Clear (2008)

showed that frozen tissue, although not perfect due to the rupture of some cells, were nevertheless suitable for staging ovaries as the oocytes, postovulatory follicles and all stages of atresia could be identified and classified. It is recommended, however, that frozen gonad material be preserved as quickly as possible to reduce the possibility of tissue deterioration while frozen. It is also recommended that any subsamples be taken from the gonad while frozen, to reduce further deterioration during the thawing process. Regardless of the method of preservation, samples should be stored with an accompanying sample label facing outwards.

Collected material should be sent to a coordinating facility (laboratory) in each country for subsampling, coordination and packing to send to a processing laboratory. If received whole from observer or port sampling activities, gonads should be sub-sampled using the approach outlined above (i.e. a core subsample (lumen to the periphery) should be taken from the mid-section of either ovary). Core subsamples should then be stored in histological cassettes in 10% buffered formalin ready for transport. Subsampling in this way will greatly reduce costs associated with sample transport, and allow the majority of the sample to remain within the CPC responsible for its collection. Otoliths and dorsal spines received from observer or port sampling activities should be cleaned, dried and placed into plastic vials and plastic bags, respectively, with an accompanying sample label, for transport and processing.

4.5 Laboratory processing, including workshops and capacity development

Ensuring standardisation and consistency in laboratory processing techniques for ageing and reproductive analyses is crucial to minimise the possibility of introducing variation and/or bias between samples as a result of variability in sample processing and storage methods between laboratories. Accordingly, it is recommended that all samples be sent to a single laboratory for independent laboratory processing. Ideally, samples should be processed in a consistent manner, and in accordance with those from the Western and Central Pacific Ocean (WCPO) to allow comparison between regions. Opportunities, in the form of workshops or training attachments, should be provided to staff from CPCs providing samples to learn current tissue processing (otolith ageing and gonad histology) and/or analytical techniques (subject to individual countries' discretion).

4.5.1 Ageing

In the laboratory, all otoliths and dorsal spines should be cleaned, dried and archived. Sectioning of otoliths and spines for annual ageing should follow protocols developed in Farley et al. (2013b; 2019). Briefly, four to five transverse sections, each approximately 300 µm thick, should be cut from each otolith, encompassing the primordium. Sections should be cleaned, dried, mounted onto glass microscope slides and covered with coverslips for reading. Each otolith section should be read independently by two readers, following the validated ageing protocol developed for South Pacific albacore tuna by Farley et al. (2013b). All readings should be conducted without knowledge of the size of fish, date of capture, or previous readings. Annotated images of each otolith section, marked with the counted opaque zones, should be captured using a Leica camera mounted to the microscope. Decimal ages should be calculated using an algorithm similar to that of Farley et al. (2013b), adapted for IO albacore tuna. Sectioning of otoliths for daily ageing should follow protocols of Williams et al. (2013).

4.5.2 Reproductive analyses

In the laboratory, if received whole, gonads should be weighed to the nearest 0.1 g and a core subsample should be taken and fixed in 10% buffered formalin. If gonads were frozen after collection, the sub-sample should be taken before the gonad is thawed. Tissue samples for histology should be

embedded in paraffin and standard histological sections prepared (i.e. thin sections (approximately 8 μ m) cut and stained with Harris' haematoxylin and eosin). Ovaries should be classified into the seven development classes used in Farley et al. (2019).

To estimate the variation in sizes of the most advanced group of oocytes, a sub-set of sections from ovaries with advanced yolked or migratory nucleus oocytes should be randomly sampled, and the diameters of a sub-sample of oocytes measured using a digital camera mounted on a stereo microscope.

Spawning frequency of females should be estimated using the postovulatory follicle method of Hunter and Macewicz (1985), recently applied to albacore tuna in the IO (Farley et al. 2014).

Batch fecundity should be estimated using the gravimetric method (Hunter et al. 1985) for females with late stage migratory nucleus or hydrated oocytes, following Farley et al. (2014). For each fish, a small sub-sample of tissue (e.g. 0.05–0.09 g; Farley et al. 2014) should be taken from the middle region of both ovary lobes, weighed (to the nearest 0.01 mg), and fixed in 10% buffered formalin. Each subsample should be teased apart under a stereo microscope and the number of migratory nucleus or hydrated oocytes counted. The number of oocytes per weight of the sub-sample should be raised to the weight of the ovary lobe to give an estimate of batch fecundity of the lobe, and estimates of two lobes summed to give an estimate of batch fecundity for the fish. Fecundity for the female population by length, age and month should be estimated as the product of batch fecundity at length and age, and spawning fraction at length and age in each month. Potential annual fecundity by length and age should be calculated by summing these across months.

4.5.3 Data management

Data (including biological data and associated metadata) should be considered property of the relevant CPC responsible for their collection and should be stored within the CPC according to their own data storage policies. Biological data and associated metadata should be sent to the laboratory where it should be stored in an appropriate database. Biological data resulting from any analysis (e.g. age estimations, reproductive data) should be sent back to the CPC of collection.

4.5.4 Sample management

Otoliths, dorsal spines, gonads and gonad subsamples should be archived. Any unused sample, the otolith and spine cross sections, and the ovary histology should remain in the archive and be available for future analysis. Access to the samples could then be evaluated by the relevant CPC in collaboration with the IOTC.

4.6 Timeline and budget

Two variations are presented regarding timelines and project budgets: Option A and Option B. In Option A, a total project duration of five years is considered, with the collection of biological material expected to take place in Years 2, 3 and 4. A preliminary analysis of resulting data, and review of the sampling program, is proposed to occur early in Year 3, following the collection and processing of biological material in Year 2. A calendar of activities for the study is provided in Table 6, and an indicative budget summary is provided in Table 7. In Option B, a four-year project duration is considered, with sample collection spanning two consecutive years (Table 8; Table 9). It should be noted that both the timeline and proposed budget are indicative – the exact timing of activities depends on the availability of funding, the selection of suitable consultants to implement the research plan, and the approval of partnership agreements between project partners.

The biggest risk to the project is considered to be failure to collect an adequate number of representative samples across space and time. The success of the study depends upon the cooperation and commitment of the fishing companies, vessel skippers and crew, observers and government scientists to obtain samples. To minimise the risk of failure, it is recommended that a project coordinator is appointed to organise the sampling with the national observer programmes, to provide training, and overall coordination of the project. It is also recommended that a fee per sample for collection of material be provided to the observers, and that fish are purchased at cost from fishing companies, to ensure access to samples. These provisions have been built into the budgets presented in Table 7 and Table 9.

4.7 Institutional arrangements

The project should be jointly managed by the IOTC through the formation of a Steering Committee. The Steering Committee will consult on various planning and implementation issues, and will report the progress of the project to the Scientific Committee of the IOTC annually throughout the course of the project. The executives of the IOTC should facilitate, when necessary, the cooperation of CPCs for the provision of samples. The Steering Committee should comprise as a minimum:

- IOTC Secretariat representative (Science Manager or representative)
- Scientific representatives for relevant CPCs (e.g. Japan, Taiwan, China)
- Project Coordinator (Project consultant(s))

The Project Coordinator and IOTC Secretariat should be responsible for informing the WPTmT and Scientific Committee on the progress and results of the project, through attendance at respective meetings. Day-to-day management and implementation of the project should be vested in the Project Coordinator.

5 Summary

Reducing uncertainty in knowledge of albacore tuna biology in the IOTC Area of Competence has been identified as a high priority issue by recent WPTmT Programs of Work (IOTC 2014, 2016, 2019c, d). To reduce this uncertainty, and to ensure sampling covers the broad spatial and temporal scales required to meet the needs of current stock assessment models, an Indian Ocean-wide study is required. This will require the implementation of an extensive biological sampling program that will be reliant upon obtaining samples from longline fleets by observers at sea. Sampling a small number of fish from every set from Japanese and Taiwanese longline vessels via observers forms the most efficient approach to ensure sampling covers a broad geographical and temporal spread.

6 Acknowledgements

We thank the large number of people who provided advice on Indian Ocean tuna fisheries and/or biological sampling of tunas, including Ashley Williams (Australian Bureau of Agricultural and Resource Economics and Sciences (ABARES)), Caroline Sanchez (SPC), Clivy Lim Shung (Ministry of Ocean Economy, Marine Resources, Fisheries and Shipping, Mauritius), Craig Proctor (CSIRO), Dan Fu (IOTC), Denham Parker (Rep. of South Africa Department of Agriculture, Forestry and Fisheries), Don Bromhead (Australian Fisheries Management Authority), Fathur Rochman (Research Institute for Tuna Fisheries, Bali, Indonesia), Francois Roupsard (SPC), Hidetada Kiyofuji (National Research Institute of Far Seas Fisheries, Japan), James Geehan (IOTC), Jiangfeng Zhu (Shanghai Ocean University), Julian Pepperell (Pepperell Research), Kevin Williams (WW Fisheries), Nathalie Bodin (Seychelles Fisheries Authority), Paul DeBruyn (IOTC), Ren-Fen Wu (Overseas Fisheries Development Council, Taiwan), Ririk Sulistyaningsih (Research Institute for Tuna Fisheries, Bali, Indonesia), Rob Campbell (CSIRO) and Sylvain Bonhommeau (IFREMER), as well as participants at the 7th Working Party on Temperate Tunas, held in Shimizu, Japan, July 2019. Laura Tremblay-Boyer assisted with translation of several national reports from French. Ashley Williams (ABARES) and Jennifer Devine (NIWA) provided constructive comments on an earlier draft of the report. Development of this report was funded by The Food and Agriculture Organization of the United Nations.

7 References

- Ahusan M, Adam MS, Ziyad A, Shifaz A, Shimal M, Jauharee R (2018). Maldives National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018_SC21_Nr16. Available at: https://www.iotc.org/documents/SC/21/NR16.
- Amir OA, Hamid ZA (2016). Tanzania National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2016. IOTC-2016-SC19-NR29. Available at: https://www.iotc.org/documents/tanzania--national-report-2016.
- Anon (1995). Report of the 6th Expert Consultation on Indian Ocean Tunas. Colombo, Sri Lanka 25–29
 September 1995. Available at:
 https://iotc.org/sites/default/files/documents/proceedings/1995/ec/IOTC-1995-EC6-R%5BEN%5D.pdf
- Anon (2018a). I.R. Iran National Report for the 21st Scientific Committee of the IOTC, 2018. IOTC-2018-SC21-R10. Available at: https://www.iotc.org/documents/SC/21/NR10.
- Anon (2018b). Japan National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. National Research Institute of Fars Seas Fisheries, Japan Fisheries Research and Education Agency and Fisheries Agency, Government of Japan. IOTC-2018-SC21_NR11_Rev1. Available at: https://www.iotc.org/documents/SC/21/NR11
- Assan C, Lucas J, Lucas V (2018). Seychelles National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018-SC21-NR22. Available at: https://www.iotc.org/documents/SC/21/NR22.
- Bard FX (1981). Le Thon Germon (*Thunnus alalunga*, Bonaterre 1788) de l'Océan Atlantique. De la dynamique des populations à la stratégie démographique. Thèse de Doctorat d'Etat. Université Pierre et Marie Curie. Paris XI. 334 p.
- Bach P, Sabarros PS, Floch L, Cauquil P, Depetris M, Duparc A, Dewals P, Medieu A, Bonhommeau S, Evano H, Huet J, Giannasi P, Marsac F, Lebranchu J (2018). UE-France Rapport national destine au Comité scientifique de la Commission des thons de l'océan Indien, 2018. In: Rapport de l'Union Européenne pour le Comité Scientifique de la CTOI de 2018 (Données 2017). IOTC-2018-SC21-NR05_-_EU_Rev1. Available at: https://www.iotc.org/documents/SC/21/NR05.
- Chacate OE, Mutombene R (2018). Mozambique National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018-SC21-NR18. Available at: https://www.iotc.org/documents/SC/21/NR18.
- Chang Y, Lee LK, Yeh SY (2016). Standardized CPUE of Indian Albacore caught by Taiwanese longliners from 1980 to 2014 with simultaneous nominal CPUE portion from observer data. IOTC-2016-WPTmT06-17. Available at: https://www.iotc.org/documents/cpue-taiwanese-albacore-longline-fishery-indian-ocean-validation-observer-data.
- Chen IC, Lee PF, Tzeng WN (2005). Distribution of albacore (*Thunnus alalunga*) in the Indian Ocean and its relation to environmental factors. Fisheries Oceanography 14: 71–80.
- Chen KS, Shimose T, Tanabe T, Chen CY, Hsu CC (2012). Age and growth of albacore *Thunnus alalunga* in the North Pacific Ocean. Journal of Fish Biology 80: 2328–2344.
- Chow S, Ushiama H (1995). Global population structure of albacore (*Thunnus alalunga*) inferred by RFLP analysis of the mitochondrial ATPase gene. Marine Biology 123: 39–45.
- Coelho R (2018). EU-Portugal National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. In: Rapport de l'Union Européenne pour le Comité Scientifique de la CTOI de 2018 (Données 2017). IOTC-2018-SC21-NR05_-_EU_Rev1. Available at: https://www.iotc.org/documents/SC/21/NR05.
- Dhurmeea Z, Zudaire I, Chassot E, Cedras M, Nikolic N, Bourjea J, West W, Appadoo C, Bodin N (2016). Reproductive biology of albacore tuna (*Thunnus alalunga*) in the Western Indian Ocean. PLoS One 11: e0168605.

- FA and OFDC (2018). Taiwan National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. Fisheries Agency, Council of Agriculture and Overseas Fisheries Development Council.
- Farley J, Clear N (2008). Albacore tuna: investigation of size monitoring, age composition, and spawning activity in the ETBF. Final Report to AFMA, Project No. 2006/826. 66 p.
- Farley JH, Williams AJ, Hoyle SD, Davies CR, Nicol SJ (2013a). Reproductive dynamics and potential annual fecundity of South Pacific albacore tuna (*Thunnus alalunga*). PLoS One 8: e60577.
- Farley JH, Williams AJ, Clear NP, Davies CR, Nicol SJ (2013b). Age estimation and validation for South Pacific albacore tuna (*Thunnus alalunga*). Journal of Fish Biology 82: 1523–1544.
- Farley JH, Hoyle SD, Eveson JP, Williams AJ, Davies CR, Nicol SJ (2014). Maturity ogives for South Pacific albacore tuna (*Thunnus alalunga*) that account for spatial and seasonal variation in the distributions of mature and immature fish. PLoS One 9: e83017.
- Farley J, Eveson P, Bonhommeau S, Dhurmeea Z, West W, Bodin N (2019). Growth of albacore tuna (*Thunnus alalunga*) in the western Indian Ocean using direct age estimates. IOTC-2019-WPTmT07DP-21_Rev1. Available at: https://www.iotc.org/documents/WPTmT/07/21.
- Geehan J, Hoyle S (2013). Review of length frequency data of the Taiwanese distant water longline fleet. IOTC-2013-WPTT15-41_Rev1. Available at: https://www.iotc.org/documents/review-length-frequency-data-taiwanchina-distant-water-longline-fleet.
- Gongona EB, Bayate DEE, Malvas SA, Tabois, BFS Jr, Ramiscal RV, Garvilles EG, Cadapan PEM, Demoos MB, Viron JG, Tanangonan I (2018). Philippines National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC–2018–SC21–NR21. Available at: https://www.iotc.org/documents/SC/21/NR21.
- Goodyear CP (2019). Modeling growth: consequences from selecting samples by size. Transactions of the American Fisheries Society 148: 528–551.
- Hewapathirana HPK, Gunawardane NDP, Ariaratne MM (2018). Sri Lanka National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC–2018–SC21–NR25. Available at: https://www.iotc.org/documents/SC/21/NR25.
- Hobsbawn PI, Patterson HM, Williams AJ (2018). Australian National Report to the Scientific Committee of the Indian Ocean Tuna Commission. Australian Bureau of Agricultural and Resource Economics and Sciences (ABARES), Canberra, ACT 2601. IOTC-2018-SC21-NR01. Available at: https://www.iotc.org/documents/SC/21/NR01.
- Hoyle SD, Sharma R, Herrera M (2014). Stock assessment of albacore tuna in the Indian Ocean for 2014 using Stock Synthesis. IOTC-2014-WPTmT05-24_Rev1. Available at: https://www.iotc.org/documents/indian-ocean-albacore-assessment.
- Hoyle SD, Yeh YM, Chang ST, Wu RF (2015). Descriptive analyses of the Taiwanese Indian Ocean longline fishery, focusing on tropical areas. IOTC–2015–WPTT17–INF09. Available at: https://www.iotc.org/documents/descriptive-analyses-taiwanese-indian-ocean-longline-fishery-focusing-tropical-areas.
- Hunter JR, Macewicz BJ (1985). Measurement of spawning frequency in multiple spawning fishes. In: Lasker T (Ed) An egg production method for estimating spawning biomass of pelagic fish: application to the northern anchovy, *Engraulis mordax*. NOAA Technical Report NMFS 36. US Department of Commerce, Springfield, VA 22161.
- Hunter JR, Lo NCH, Leong RJH (1985). Batch fecundity in multiple spawning fishes. In: Lasker T (Ed) An egg production method for estimating spawning biomass of pelagic fish: application to the northern anchovy, *Engraulis mordax*. NOAA Technical Report NMFS 36. US Department of Commerce, Springfield, VA 22161.
- IEO and SGP (2018). EU-SPAIN National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. In: Rapport de l'Union Européenne pour le Comité Scientifique de la CTOI de

- 2018 (Données 2017). IOTC-2018-SC21-NR05_-_EU_Rev1. Available at: https://www.iotc.org/documents/SC/21/NR05.
- IOTC (2014). Report of the fifth session of the IOTC Working Party on Temperate Tunas. IOTC-2014-WPTmT05-R. Available at: https://www.iotc.org/documents/report-5th-session-working-party-temperate-tunas.
- IOTC (2016). Report on the sixth session of the IOTC Working Party on Temperate Tunas. IOTC–2016—WPTmT06–R[e]. Available at: https://www.iotc.org/documents/report-sixth-working-party-temperate-tunas.
- IOTC (2019a). Catch and effort data all vessels. IOTC-2019-WPTmT07-DATA07. Available at: https://www.iotc.org/WPTmTa/07/Data/07-CEAII.
- IOTC (2019b). Files for albacore stock assessment. IOTC-2019-WPTmT07-DATA12. Available at: https://www.iotc.org/WPTmTa/07/Data/12-SA.
- IOTC (2019c). Revision of the WPTmT program of work. IOTC–2019–WPTmT07(DP)–08. Available at: https://www.iotc.org/documents/WPTmT/07/08.
- IOTC (2019d). Report of the seventh session of the IOTC Working Party on Temperate Tunas: Assessment Meeting. IOTC-2019-WPTmT07(AS)-R[E]. Available at: https://www.iotc.org/documents/WPTmT/702/R E.
- IOTC (2019e). Report on the seventh session of the IOTC Working Party on Temperate Tunas (Data Preparatory Session). IOTC-2019-WPTmT07(DP)-R[E]. Available at: https://www.iotc.org/documents/WPTmT/07/ReportDP E.
- Khan MF (2018). Pakistan's National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018-SC21-NR20. Available at: https://www.iotc.org/documents/SC/21/NR20.
- Kim DN, Lee SI, Lee MK, An DH (2018). Korea National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC–2018–SC21–NR13. Available at: https://www.iotc.org/documents/SC/21/NR13.
- Kitakado T, Fiorellato F, De Bruyn P (2019). Allometric curve for the Indian Ocean albacore. IOTC-2019-WPTmT07AS-INFO2. Available at: https://www.iotc.org/documents/WPTmT/702/INFO2.
- Langley A (2019). Stock assessment of albacore tuna in the Indian Ocean using Stock Synthesis for 2019. IOTC-2019-WPTmT07(AS)-11. Available at: https://www.iotc.org/documents/WPTmT/702/11.
- Langley A, Hoyle S (2016). Stock assessment of albacore tuna in the Indian Ocean using Stock Synthesis. IOTC–2016–WPTmT06–25. Available at: https://www.iotc.org/documents/albacore-stock-assessment-using-ss.
- Liorzou B (1989). Les nouveaux engins de pêche pour la capture du germon. Description, statistiques, impact sur le stock Nord-Atlantique. Collective Volumes of Scientific Paper ICCAT 30(1); 203–217.
- Lirdwitayaprasit P, Wongkeaw A, Yothakong T (2018). Thailand National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018-SC21-NR29. Available at: https://www.iotc.org/documents/SC/21/NR29.
- MAF (2018). Ministry of Agriculture and Fisheries, Sultanate of Oman National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018-SC21-NR19. Available at: https://www.iotc.org/documents/SC/21/NR19.
- Matsumoto T (2016). Review of Japanese longline fishery and its albacore catch in the Indian Ocean. IOTC-2016-WPTmT06-14. Available at: https://www.iotc.org/documents/review-japanese-longline-fishery-and-its-albacore-catch-indian-ocean-2.
- Methot RD Jr, Wentzel, CR (2013). Stock synthesis: a biological and statistical framework for fish stock assessment and fishery management. Fisheries Research 142: 86–99.

- Montes I, Iriondo M, Manzano C, Arrizabalaga H, Jiménez E, Pardo MA, Goñi N, Davies CA, Estonba A (2012). Worldwide genetic structure of albacore *Thunnus alalunga* revealed by microsatellite DNA markers. Marine Ecology Progress Series 471: 183–191.
- MRHP, CSP, USTA (2018). Rapport National de Madagascar destiné au Comité Scientifique de la Commission des thons de l'Ocean Indien, 2018. IOTC-2017-SC21-NR14. Ministère des Ressources Halieutiques et de la pêche (MRHP), Centre de Surveillance de Pêche (CSP and Unité Statistique Thonière d'Antsiranana (USTA). Available at: https://www.iotc.org/documents/SC/21/NR14.
- Mueni E, Ndegwa S, Okemwa G (2018). [KENYA] National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC–2018–SC21–NR12. Available at: https://www.iotc.org/documents/SC/21/NR12.
- Nikolic N, Morandeau G, Hoarau L, West W, Arrizabalaga H, Hoyle S, Nicol SJ, Bourjea J, Puech A, Farley JH, Williams AJ, Fonteneau A (2017). Review of albacore tuna, *Thunnus alalunga*, biology, fisheries and management. Reviews in Fish Biology and Fisheries 27: 775–810.
- Otsu T, Uchida RN (1959). Sexual maturity and spawning of albacore in the Pacific Ocean. Fishery Bulletin 148: 287–305.
- Panjarat S, Rodpradit S, Singtongyam W (2016). Albacore tuna unloaded at Phuket Fishing Port, Thailand. IOTC-2016-WPTmT06-10. Available at: https://www.iotc.org/documents/albacore-tuna-unloaded-phuket-fishing-port-thailand.
- Parker D, Winker H, da Silva C, Mketsu Q, Kerwath S (2018). South African National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC–2018–SC21–NR26. Available at: https://www.iotc.org/documents/SC/21/NR26.
- Premchand, Ramalingam L, Tiburtius A, Siva A, Das A, Sanadi RB, Tailor RKB (2015). India's National Report to the Scientific Committee of the Indian Ocean Tuna Commission 2015. IOTC–2015–SC18—NR09[E]. Available at: https://www.iotc.org/documents/india—national-report-2015.
- Ruchimat T, Fahmi Z, Setyadji B, Yunanda T (2018). Indonesia National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018-SC21-NR09. Available at: https://www.iotc.org/documents/SC/21/NR09.
- Samsudin B, Sallehudin J, Tengku, BTS, Nor Azlin M (2018). Malaysia National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018-SC21-NR15. Available at: https://www.iotc.org/documents/SC/21/NR15.
- Sheik Mamode S, Lim Shung C, Mootoosamy L (2018). Mauritius National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC–2018–SC21–NR17. Available at: https://www.iotc.org/documents/SC/21/NR17.
- Skeik Heile AI (2018). Somali National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018-SC21-NR24. Available at: https://www.iotc.org/documents/SC/21/NR24.
- SPC-OFP (2018). Estimates of annual catches in the WCPFC statistical area. Information paper WCPFC-SC14-2018 ST-IP-01 presented to the fourteenth regular session of the Scientific Committee of the Western and Central Pacific Fisheries Commission, Busan, Republic of Korea.
- UE-Italy (2018). UE-Italy National Report to the Scientific Committee of the Indian Ocean Tuna Commission (IOTC), 2017. In: Rapport de l'Union Européenne pour le Comité Scientifique de la CTOI de 2018 (Données 2017). IOTC-2018-SC21-NR05_-_EU_Rev1. Available at: https://www.iotc.org/documents/SC/21/NR05.
- Ueyanagi S (1969). Observations on the distribution of tuna larvae in the Indo-Pacific Ocean with emphasis on the delineation of the spawning areas of albacore, *Thunnus alalunga*. Far Seas Research Laboratories Bulletin 2: 177–256.
- United Kingdom (2018). United Kingdom (EU) IOTC Scientific Committee Report 2017. In: Rapport de l'Union Européenne pour le Comité Scientifique de la CTOI de 2018 (Données 2017). IOTC-2018-SC21-NR05 EU_Rev1. Available at: https://www.iotc.org/documents/SC/21/NR05.

- Wang SP (2019). Data analysis and CPUE standardization of albacore caught by Taiwanese longline fisheries in the Indian Ocean. IOTC-2019-WPTmT07(DP)-14_Rev1. Available at: https://www.iotc.org/documents/WPTmT/08/14.
- Williams AJ, Farley JH, Hoyle SD, Davies CR, Nicol SJ (2012). Spatial and sex-specific variation in growth of albacore tuna (*Thunnus alalunga*) across the South Pacific Ocean. PLoS One 7: e39318.
- Williams AJ, Leroy BM, Nicol SJ, Farley JH, Clear NP, Krusic-Golub K, Davies CR (2013). Comparison of daily- and annual-increment counts in otoliths of bigeye (*Thunnus obesus*), yellowfin (*T. albacares*), southern bluefin (*T. maccoyii*) and albacore (*T. alalunga*) tuna. ICES Journal of Marine Science 70: 1439–1450.
- Xu Y, Sippel T, Teo SLH, Piner K, Chen KS, Wells RJ (2014). A comparison study of North Pacific albacore (*Thunnus alalunga*) age and growth among various sources. ISC/14/ALBWG/04 ISC Albacore Working Group Meeting, 14–28 April 2014, La Jolla, CA, 92037, USA.
- Yeh SY, Hui CF, Treng TD, Kuo CL (1996). Indian Ocean albacore stock structure studies by morphometric and DNA sequence methods. In: Anganuzzi AA, Stobberup KA, Webb NJ (eds) Proceedings of the 6th Expert Consultation on Indian Ocean Tunas.
- Zhu J, Wu F, Yang X (2018). [China] National Report to the Scientific Committee of the Indian Ocean Tuna Commission, 2018. IOTC-2018-SC21-NR02. Available at: https://www.iotc.org/documents/SC/21/NR02.
- Zou L, Dai X, Zhu J (2014). Size composition of Indian Ocean albacore based on Chinese longline fishery. IOTC-2014-WPTmT05-14. Available at: https://www.iotc.org/documents/size-composition-indian-ocean-albacore-based-chinese-longline-fishery.

Table 1: Parameters evaluated and a description of the model sensitivities investigating the influence of key biological parameters from the base Indian Ocean albacore tuna assessment model of Langley (2019).

Model	Base model parameters (Langley 2019)	Alternative parameters examined (this study)	Description of alternative parameters
	, , ,	, , ,	•
LengthAtAge	CVs for length-at-age	CV_young 0.10	Increase variation in length-
	CV_young 0.06 CV_old 0.025	CV_old 0.10	at-age for males and females.
LengthWeight	<i>α</i> = 1.3718e-05	<i>a</i> = 6.9e-06	Use length-weight parameters
	b = 3.0973	<i>b</i> = 3.2263	from Kitakado et al. (2019)
GrowthYoung	Female $k = 0.38$	Female <i>k</i> = 0.478	Investigate potential bias in
	Female Length1= 52.6	Female Length1= 39.4	growth for youngest age
	Female <i>Linf</i> = 103.8	Female <i>Linf</i> = 103.8	classes by constraining VB
	Male $k = 0.34$	Male <i>k</i> = 0.422	growth functions to pass
	Male Length1= 52.0	Male <i>Length1</i> = 38.05	through origin (Age 0, Length
	Male <i>Linf</i> = 110.6	Male <i>Linf</i> = 110.6	0).
<i>MaturityOgive</i>	Length based	Length based maturity	Use maturity at length from
	Reproductive Potential	ogive from Dhurmeea et	recent western Indian Ocean
	from South Pacific	al. (2016)	study (rather than South
	albacore tuna.		Pacific reproductive potential
			at length)
NatMortAge	M = 0.30 all ages	Females	Increase natural mortality of
		M = 0.30 Ages 1-3	females aged 4+ yr from 0.30
		M = 0.35 Ages >= 4	to 0.35
EastWest			Two region model,
			recruitment variation in both
			east and west regions.
			No movement W<>E
EastWestGrowth	Female <i>k</i> = 0.38	FemaleW <i>k</i> = 0.38	Differences in growth
	Male $k = 0.34$	MaleW $k = 0.34$	between east and west IO.
		FemaleE <i>k</i> = 0.532	Higher (+40%) growth rate (k)
		MaleE <i>k</i> = 0.476	in East relative to W (base values).
			No movement W<>E
			NO MOVEMENT VV V/L

Table 2: Change in the values of the likelihood components relative to the Base model. A deterioration (increase in LL) in the likelihood (of at least 1.0) is highlighted in red. An improvement in the likelihood (of at least 1.0) is highlighted in green. The two region models are compared separately.

Model	Total	CPUE	LF	Recruit	Priors
BASE	881.36	-139.79	1053.81	-32.76	0.00
Growth Young	-53.09	-4.52	-48.17	0.16	-0.55
LengthAtAge	1.07	-7.05	8.77	-0.73	0.00
LengthWeight	-0.33	-0.06	-0.29	0.02	0.00
Mature1	0.06	-0.13	0.24	-0.04	0.00
NatMortAge	1.73	-0.11	1.85	0.00	0.00
EastWest	758.07	-262.80	1049.04	-33.71	0.00
EastWestGrowth	-53.96	6.97	-59.67	0.43	-0.07

Table 3: Changes in individual components relative to the Base model. A deterioration in the likelihood (of at least 1.0) is highlighted in red. An improvement in the likelihood (of at least 1.0) is highlighted in green. The two region models are compared separately.

Model	CPUE Length co					
	LL3	LL4	LL1	LL2	LL3	LL4
BASE	-134.82	-	258.70	177.04	279.13	243.92
Growth Young	-4.88	-	-5.83	-5.15	-16.96	-2.40
LengthAtAge	-7.00	-	-2.37	1.39	21.30	21.49
LengthWeight	-0.05	-	-0.05	0.00	-0.01	0.00
MaturityOgive	-0.13	-	-0.03	0.02	-0.12	-0.05
NatMortAge	-0.12	-	0.83	0.57	0.25	0.08
EastWest	-136.99	-120.42	256.14	175.95	276.32	240.21
EastWestGrowth	-0.62	7.68	-1.09	-10.77	0.77	-33.68

Table 4: Comparison of stock status and yields and associated standard deviations (derived from covariance matrix) from the range of model options. Presented are absolute values (upper) and percentage difference from the base model (lower) (>10% reduction highlighted in red, >10% increase highlighted in green).

Model		SBO SB2017		SB20:	17/SBMSY	F20	017/FMSY		MSY	
	Value	SD	Value	SD	Value	SD	Value	SD	Value	SD
BASE	103,612	6,265	26,567	6,290	1.246	0.226	1.227	0.202	33,302	1,854
Growth Young	110,327	4,590	23,419	5,079	0.994	0.179	1.469	0.222	31,333	1,079
LengthAtAge	91,320	2,941	15,816	3,194	0.827	0.146	1.676	0.226	30,132	745
LengthWeight	106,166	6,468	27,322	6,493	1.240	0.225	1.230	0.204	33,252	1,848
MaturityOgive	130,753	8,531	37,031	8,286	1.311	0.217	1.132	0.187	34,266	2,012
NatMortAge	84,689	5,278	22,673	5,331	1.283	0.230	1.200	0.198	33,748	1,928
EastWest	123,935	-	51,318	-	1.964	-	0.636	-	38,935	0
EastWestGrowth	129,299	-	54,739	-	1.967	-	0.667	-	39,679	0
Growth Young	6%	-27%	-12%	-19%	-20%	-21%	20%	10%	-6%	-42%
LengthAtAge	-12%	-53%	-40%	-49%	-34%	-35%	37%	12%	-10%	-60%
LengthWeight	2%	3%	3%	3%	0%	0%	0%	1%	0%	0%
MaturityOgive	26%	36%	39%	32%	5%	-4%	-8%	-8%	3%	9%
NatMortAge	-18%	-16%	-15%	-15%	3%	2%	-2%	-2%	1%	4%
									.=	
EastWest	20%		93%		58%		-48%		17%	
EastWestGrowth	25%		106%		58%		-46%		19%	

Table 5: Estimated number of hooks and sets deployed, catches and potential sampling yield per assessment region for the Taiwanese longline fleet. The estimated number of fish available for sampling is based on a uniform observer coverage of 2% of sets per region and a sampling strategy whereby a maximum of five albacore tuna (i.e., the first five encountered) are sampled in each set.

Region	Total no. hooks deployed ¹	Estimated no. sets ²	Catch (no.) ¹	% zero catches ³	Number of fish available for sampling ⁴
1	72,997,562	20,856	209,487	90	209
2	7,768,200	2,219	2,258	80	44
3	50,859,807	14,531	613,487	20	1,163
4	9,379,958	2,680	189,645	20	214

¹ Source: IOTC 2019a

² Assuming an average of 3,500 hooks per set (Hoyle et al. 2015)

³ Approximated for albacore tuna assessment regions from Hoyle et al. (2015)

⁴ Assuming 2% of sets are observed and 5 fish are sampled from each set

Table 6: Timeline of proposed project activities under Option A.

Activity/Milestone	Year 1	Ye	ar 2	Yea	ar 3	Yea	ar 4	Yea	ar 5
IOTC Scientific Committee endorsement	Х								
IOTC Secretariat approval	Χ								
Selection of Project Coordinator (lead consultant)	Χ								
Signing of contracts	Х								
Negotiate sampling arrangements and obtain endorsement/support	Х	Х							
Training in sample collection		Χ							
Collection of biological material		Χ	Χ	Χ	Χ	Χ	Χ		
Laboratory analysis			Χ	Χ	Χ	Χ	Χ	Χ	
Preliminary data analysis and program review				Χ					
Data analysis								Χ	Χ
SC reporting									Х
Project report and recommendations									Х

Table 7: Indicative budget (thousand USD) for the observer-based sampling, processing, analyses and reporting on Indian Ocean albacore biology under Option A.

Note: Institutional overheads of consulting organisations are not included in this estimated budget.

Item	Year 1	Year 2	Year 3	Year 4	Year 5
Project coordination ^a	30	30	30	30	30
Travel (for project coordination, observer training)	20	10	10		
Sampling equipment	5	5	5	3	
Costs for purchasing fish (US 60 per sample) ^b		120	120	120	
Reward payment for observers (USD 20 per sample)		30	30	30	
Transportation of biological samples		10	20	20	
Processing of otoliths for annual ageing ^c		80	80	80	
Processing of otoliths and dorsal fin spines for daily ageing ^d			5	2	
Processing of gonads for reproductive analyses ^e		40	40	40	
Data analysis and reporting ^a			40	20	50
Travel (to WPTmT/Scientific Committee meetings)		10	15	10	15
Miscellaneous costs (e.g., development of training materials)	20	5	5	5	5
Total	75	350	410	370	100

^a Includes daily rates of USD 500 per day

e Assumes a cost of USD 40 per sample and a sex ratio of 1:1 (i.e., 1000 females sampled in a given year)

^b Assumes a cost of USD 5 per kg, and an average fish weight of 12 kg

^c Assumes a cost of USD 40 per sample, and 2000 fish sampled in each year

^d Assumes a cost of USD 60 per sample

Table 8: Timeline of proposed project activities under Option B.

Activity/Milestone		Year 1		Year 2		Year 3		ar 4
IOTC Scientific Committee endorsement	Χ							
IOTC Secretariat approval	Χ							
Selection of Project Coordinator (lead consultant)		Χ						
Signing of contracts		Χ						
Negotiate sampling arrangements and obtain endorsement/support		X	Х					
Training in sample collection			Χ					
Collection of biological material			Χ	Χ	Χ	Χ		
Laboratory analysis				Χ	Χ	Χ	Χ	Χ
Preliminary data analysis and program review					Χ			
Data analysis							Χ	Χ
SC reporting								Χ
Project report and recommendations								Χ

Table 9: Indicative budget (thousand USD) for the observer-based sampling, processing, analyses and reporting on Indian Ocean albacore biology under Option B. Note: Institutional overheads of consulting organisations are not included in this estimated budget.

Item	Year 1	Year 2	Year 3	Year 4	
Project coordination ^a	30	30	30	30	
Travel (for project coordination, observer training)	20	10	10		
Sampling equipment	5	5	5		
Costs for purchasing fish (US 60 per sample) ^b		120	120		
Reward payment for observers (USD 20 per sample)		40	40		
Transportation of biological samples		10	20		
Processing of otoliths for annual ageing ^c		80	80		
Processing of otoliths and dorsal fin spines for daily ageing ^d			2		
Processing of gonads for reproductive analyses ^e		40	40		
Data analysis and reporting ^a			20	45	
Travel (to WPTmT/Scientific Committee meetings)		10	15	10	
Miscellaneous costs (e.g., development of training materials)	20	5	5	5	
Total	75	350	387	90	

^a Includes daily rates of USD 500 per day

^b Assumes a cost of USD 5 per kg, and an average fish weight of 12 kg

^c Assumes a cost of USD 40 per sample, and 2000 fish sampled in each year

^d Assumes a cost of USD 60 per sample

^e Assumes a cost of USD 40 per sample and a sex ratio of 1:1 (i.e., 1000 females sampled in a given year)

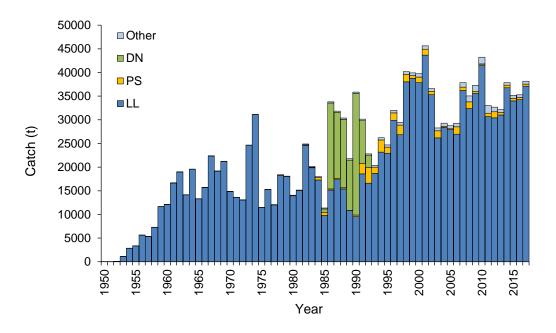


Figure 1: Total annual catch (mt) of albacore tuna by fishing method from 1950 to 2017. DN, driftnet; PS, purse-seine; LL, longline. Source: IOTC 2019b.

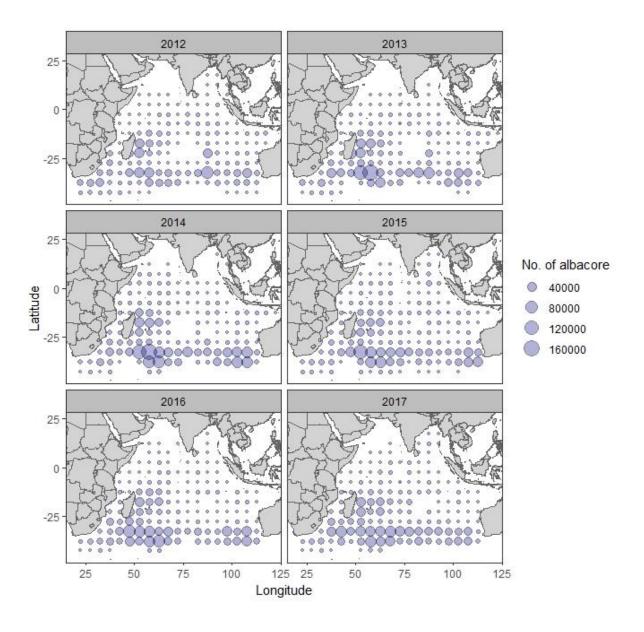


Figure 2: Total annual longline catches of albacore tuna, 2012–2017. The blue circles represent the aggregated longline catch (numbers of fish, all fleets) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch.

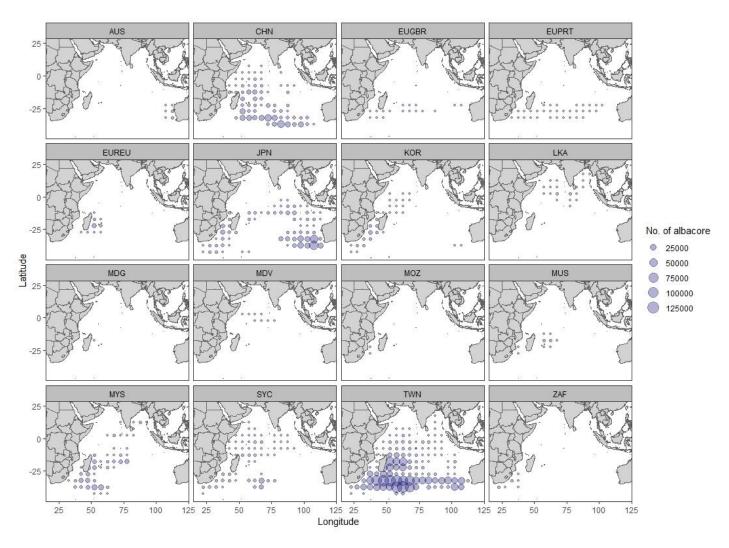


Figure 3: Total longline catch of albacore tuna by fleet, 2016–2017. The blue circles represent the aggregated longline catch (numbers of fish) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch. AUS = Australia, CHN = China, EUGBR = United Kingdom, EUPRT = Portugal, EUREU = France-Reunion, JPN = Japan, KOR = Republic of Korea, LKA = Sri Lanka, MDG = Madagascar, MDV = Maldives, MOZ = Mozambique, MUS = Mauritius, MYS = Malaysia, SYC = Seychelles, TWN = Taiwan, China, ZAF = Republic of South Africa.

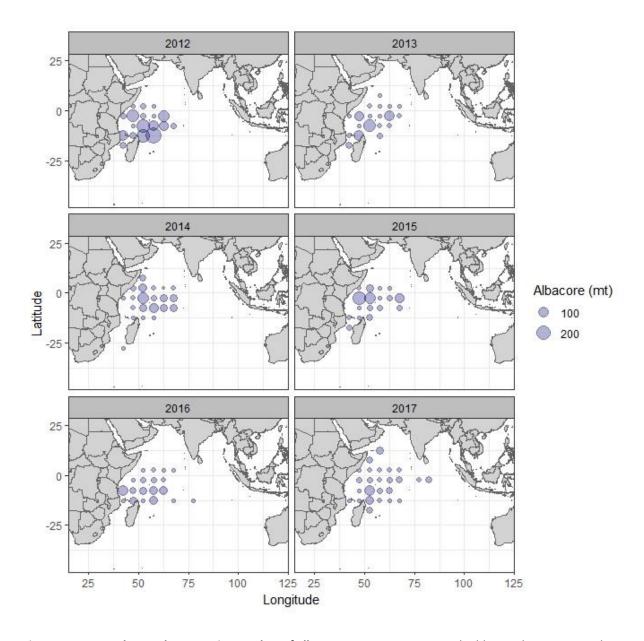


Figure 4: Total annual purse-seine catches of albacore tuna, 2012–2017. The blue circles represent the aggregated catch (weight of fish in mt, all fleets) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch.

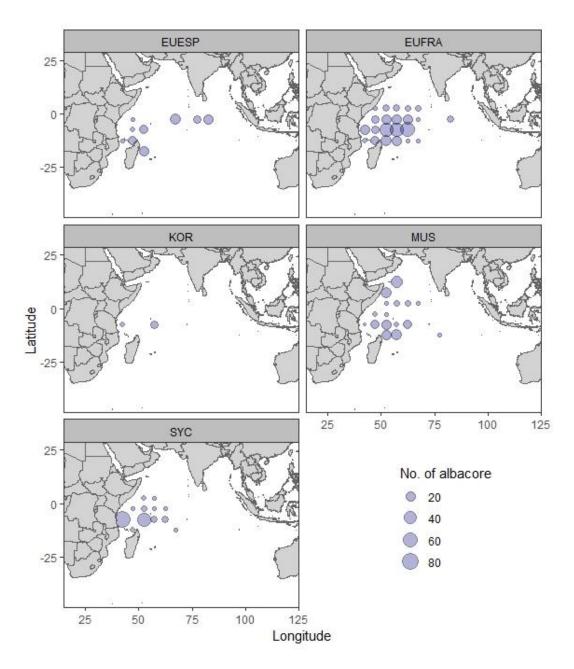


Figure 5: Total purse-seine catch of albacore tuna by fleet, 2016–2017. The blue circles represent the aggregated catch (numbers of fish) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch.

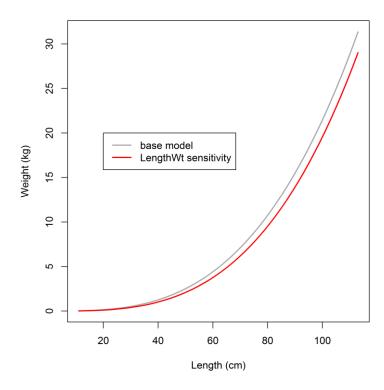


Figure 6: Comparison of the length-weight relationship used in the base model and the model sensitivity LengthWeight.

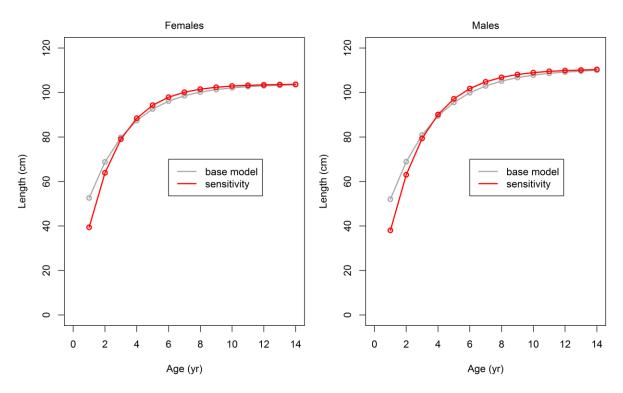


Figure 7: Comparison of the growth functions used in the base model and the model sensitivity *GrowthYoung*.

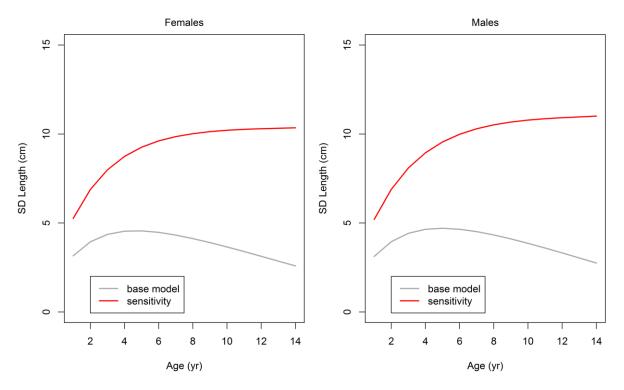


Figure 8: Parameterisation of the variation in length-at-age from the base model and the model sensitivity *LengthAtAge*.

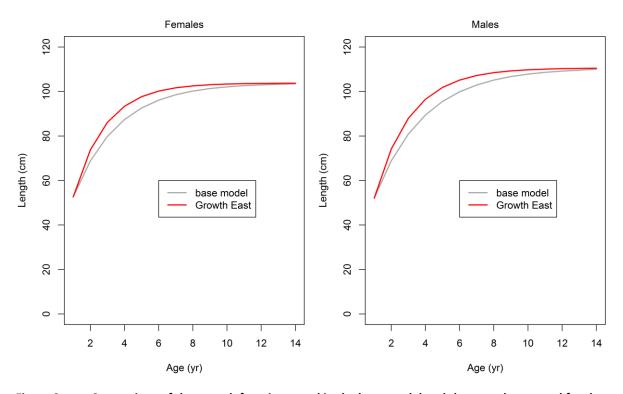


Figure 9: Comparison of the growth functions used in the base model and the growth assumed for the eastern Indian Ocean in the *EastWestGrowth* model sensitivity. Growth in the western Indian Ocean is assumed to be equivalent to the growth in the base model.

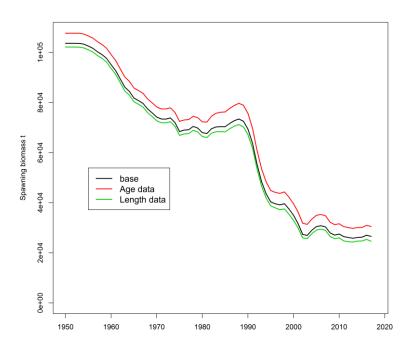


Figure 10: Spawning biomass trajectories for the base model and the base model with simulated length composition data or simulated age composition data included.

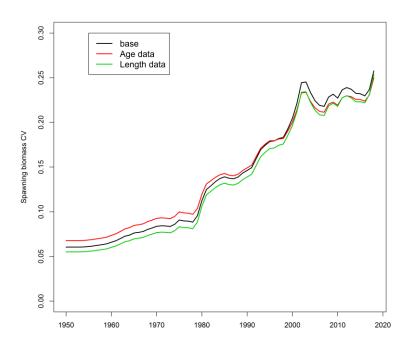


Figure 11: Annual CV of the estimated spawning biomass for the base model and the base model with simulated length or age composition data included.

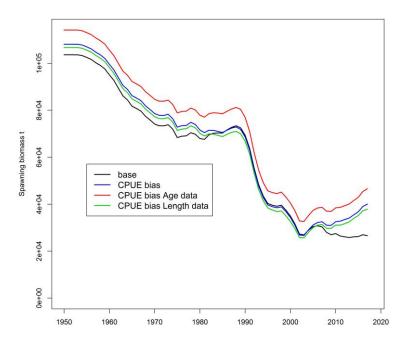


Figure 12: Spawning biomass trajectories for the base model, the biased CPUE model and the biased CPUE model with simulated length or age composition data included.

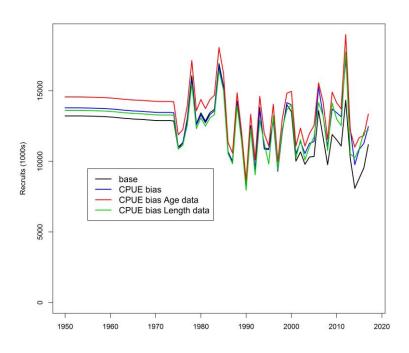


Figure 13: Annual recruitment for the base model, the biased CPUE model and the biased CPUE model with simulated length or age composition data included.

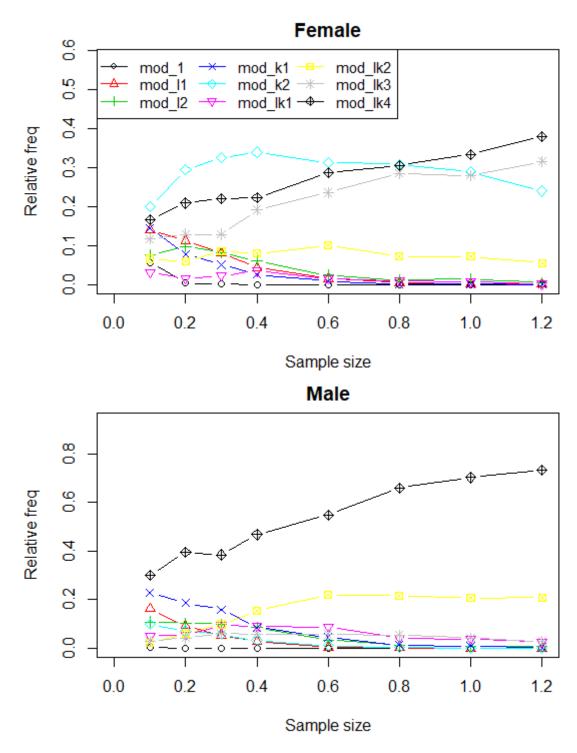


Figure 14: Frequency of growth model selection given differing sample sizes for female (top) and male (bottom) albacore tuna. Models are described in the text. Results are based on bootstrap resampling of Pacific length at age data at different proportions of the true sample sizes.

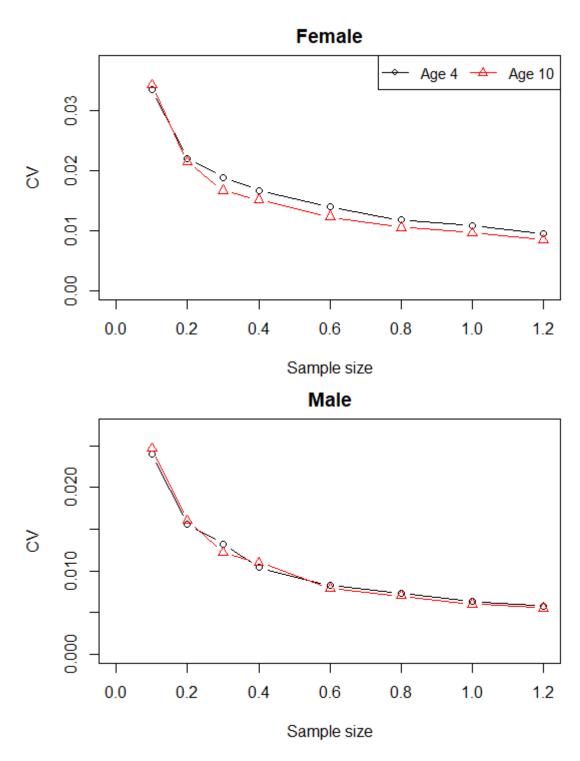


Figure 15: CV of predicted size at ages 4 and 10 for female (top) and male (bottom) albacore tuna. Results are based on bootstrap resampling of Pacific length at age data at different proportions of the true sample sizes.

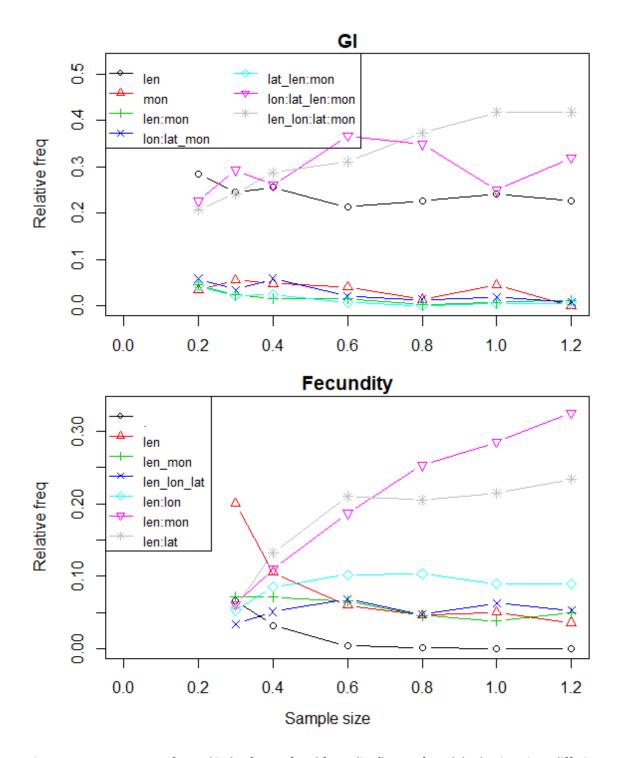


Figure 16: Frequency of gonad index (GI, top) and fecundity (bottom) model selection given differing sample sizes for albacore tuna. Models are described in the text. Results are based on bootstrap resampling of Pacific GI and fecundity data at different proportions of the true sample sizes. Rarely selected models are omitted to avoid cluttering the figures.

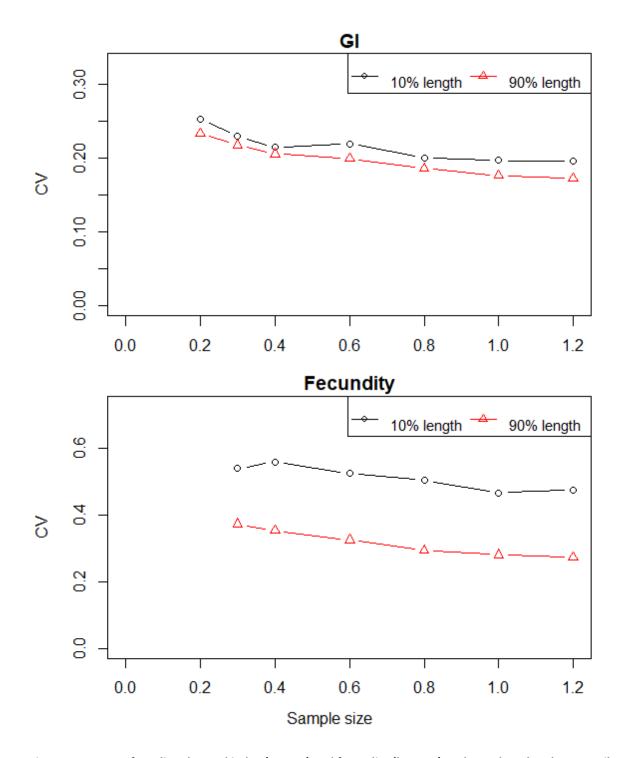


Figure 17: CV of predicted gonad index (GI, top) and fecundity (bottom) at the 10th and 90th percentiles of length for albacore tuna. Results are based on bootstrap resampling of Pacific GI and fecundity data at different proportions of the true sample sizes.

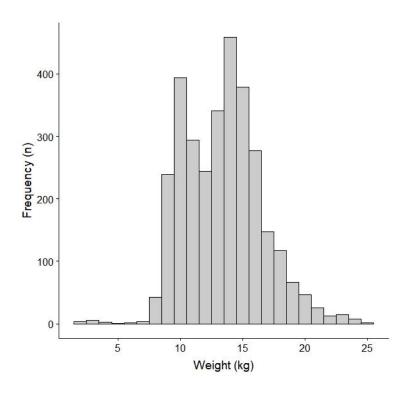


Figure 18: Weight frequency of individual albacore tuna measured from the Western Tuna and Billfish Fishery port sampling program, Australia. Data are for fish collected between July 2015 and June 2018 (Source: AFMA).

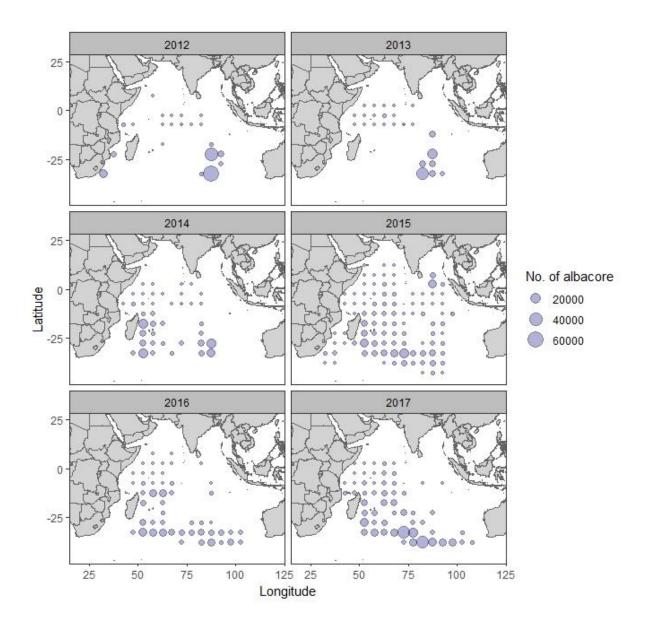


Figure 19: Total annual catch of albacore tuna by Chinese-flagged longline vessels, 2012–2017. The blue circles represent the aggregated longline catch (numbers of fish) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch.

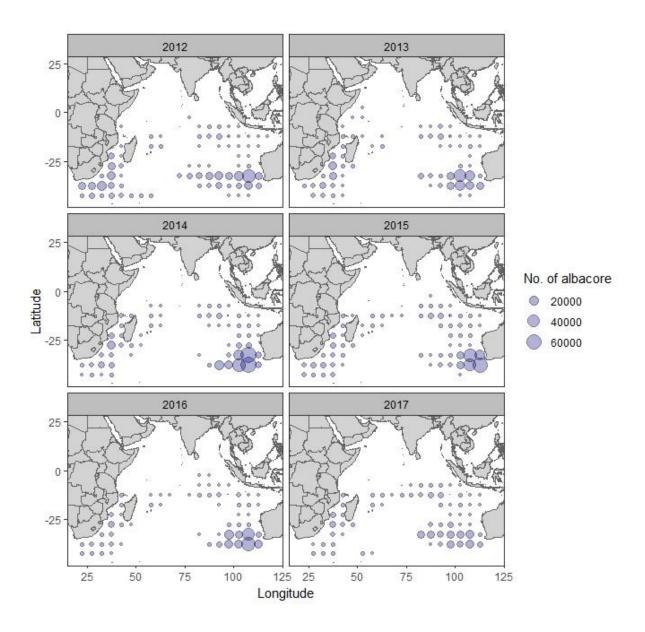


Figure 20: Total annual catch of albacore tuna by Japanese-flagged longline vessels, 2012–2017. The blue circles represent the aggregated longline catch (numbers of fish) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch.

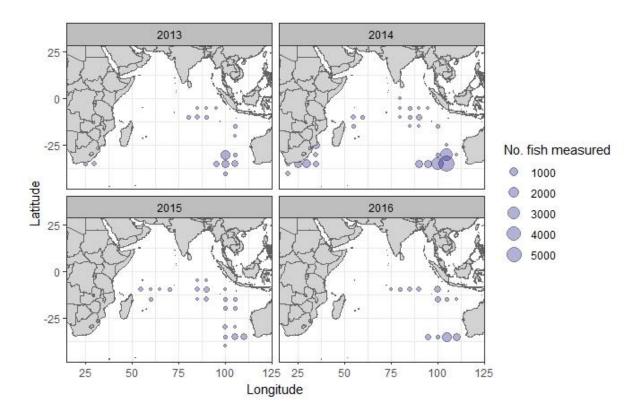


Figure 21: Numbers of albacore tuna measured by observers on Japanese longline vessels, 2013–2016. Source: IOTC 2019e. Note data for 2017 and 2018 were not available.

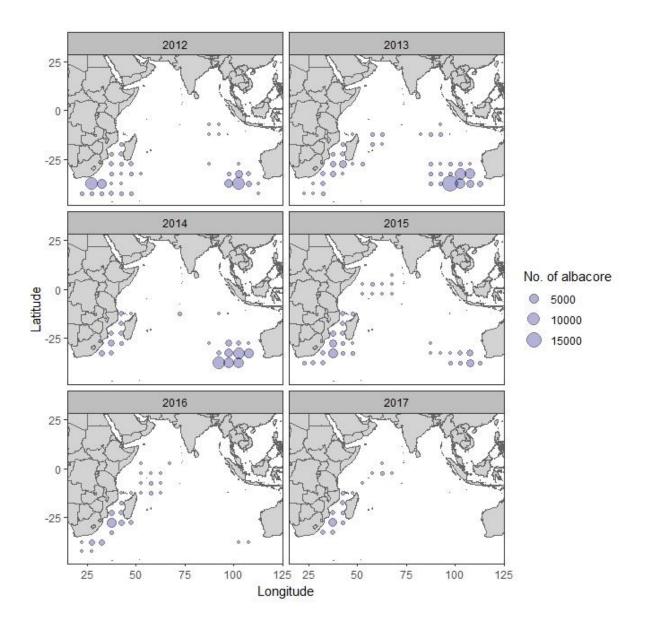


Figure 22: Total annual catch of albacore tuna by Korean-flagged longline vessels, 2012–2017. The blue circles represent the aggregated longline catch (numbers of fish) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch.

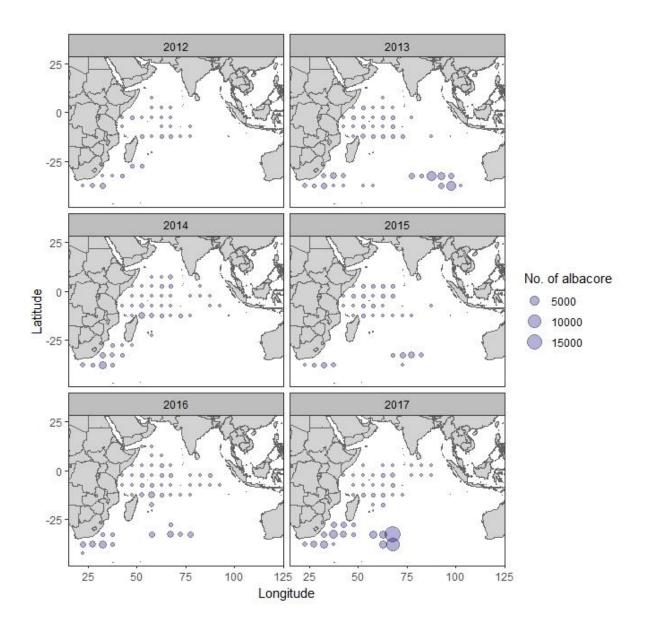


Figure 23: Total annual catch of albacore tuna by Seychelles-flagged longline vessels, 2012–2017. The blue circles represent the aggregated longline catch (numbers of fish) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch.

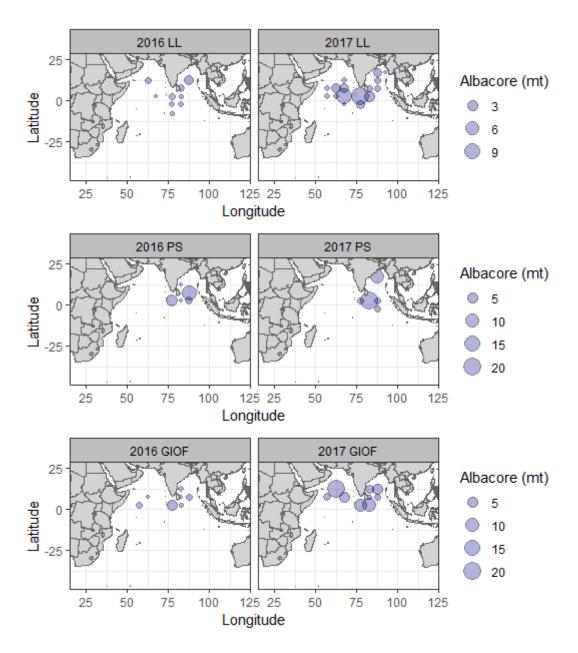


Figure 24: Total annual catch of albacore tuna by Sri Lankan-flagged vessels, 2016 and 2017. The blue circles represent the aggregated longline catch (numbers of fish) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch. LL = longline, PS = purse-seine, GIOF = gillnet (offshore).

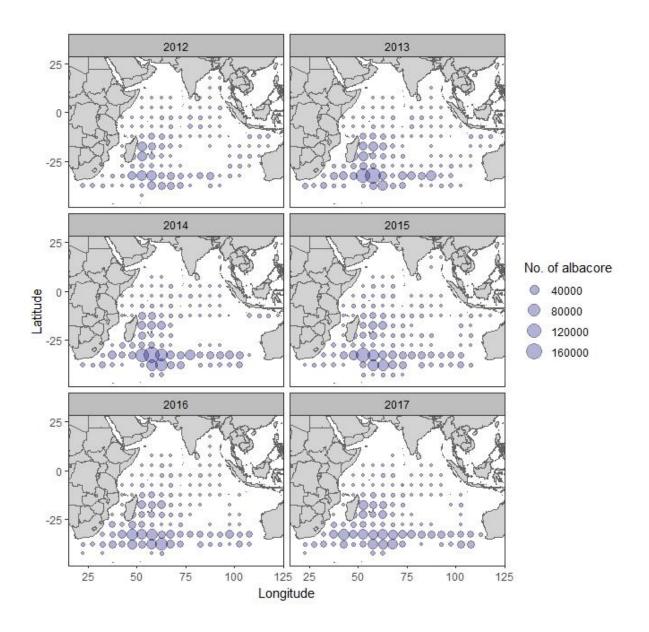


Figure 25: Total annual catch of albacore tuna by Taiwanese-flagged longline vessels, 2012–2017. The blue circles represent the aggregated longline catch (numbers of fish) by 5-degree cell. The area of the circles is proportional to the magnitude of the catch.